

Studies on Bio-denitrification of Wastewater Using Immobilized GAC in Draft Tube Spouted Bed Reactor

Keshava Joshi¹, Lokeshwari Navalgund^{1*}, Vinayaka B. Shet²

¹ Department of Chemical Engineering, SDM College of Engineering and Technology, (Visvesvaraya Technological University, Belagavi), Dharwad, Karnataka, 580 002, India.

² Department of Biotechnology Engineering, NMAM Institute of Technology-Affiliated to NITTE (Deemed to be University), Nitte-574110, Karnataka, India.

ARTICLE INFO

ORIGINAL ARTICLE

Article History:

Received: 15 September 2024

Accepted: 20 November 2024

*Corresponding Author:

Lokeshwari Navalgund

Email:

lokeshwarinavalgund@gmail.com

Tel:

+83 62328207

Keywords:

Biomass,

Biofilm,

Bioreactor,

Granular activated carbon,

Nitrate,

Pseudomonas syringae.

ABSTRACT

Introduction: The draft tube-spouted bed bioreactor with GAC particles immobilized with *Pseudomonas syringae* is being evaluated to study the effect of suspended biomass and biofilm thickness on the rate of denitrification. Though the biofilm thickness is not directly controlled in wastewater treatment by the diffusion limitation and consequent substrate penetration in the biofilm, biofilm thickness will probably have a significant impact on bacteriological activity.

Materials and Methods: The reactor studies were accomplished to study the result of dilution rate on attached biomass, suspended biomass, and biofilm thickness with nitrate reduction under steady-state conditions. A spouted bed reactor with the growth media prepared was used to study the bio-denitrification using *Pseudomonas syringae*.

Results: The study of the attached biomass on nitrate reduction indicated that, as the attached biomass increased from 0.35 g/g to 0.54 g/g at a 0.166/h dilution rate, the nitrate reduction percentage decreased from 98.18% to 88.2%. During the study, it was observed that the biomass and biofilm thickness increased and decreased, respectively, with a rise in influent nitrate concentration and dilution rate. The rise in dilution rate as well as influent nitrate concentration throughout the study increased the rate of suspended biomass.

Conclusion: The nitrate reduction rate was high with higher rates of loading in a draft tube spouted bed bioreactor, due to well-organized recirculation of the solids inside the reactor. The formation of biofilm thickness on solids is a significant character as it increases the nitrate reduction rate to meet the effluent standards.

Citation: Joshi K, Navalgund L, B. Shet V. *Studies on Bio-denitrification of Wastewater Using Immobilized GAC in Draft Tube Spouted Bed Reactor*. J Environ Health Sustain Dev. 2024; 9(4): 2470-9.

Introduction

Protecting the aquatic environment has grown to be a significant global concern because of population growth and industrialization¹. High nitrate and ammonia concentration with lower DO levels have an impact on aquatic life, human life and drinking water issues. These are the few negative consequences that can result from the build-up of different types of nitrogen and its

different forms in the different water bodies. Electrodialysis, Ozonation, Reverse osmosis, are the physical and chemical processes utilized to reduce nitrate^{2, 3}. However, the high capital and energy costs limit the application of these treatments⁴. Biological nitrate removal is the most economical strategy and enhanced option to use because it does not produce any by-products⁵.

Owing to their increased efficiency per unit

biomass, bioreactors with immobilized bacteria are perfect for laboratories and small firms with limited area for a traditional treatment site⁶. According to study investigations, immobilized attached cells on a diverse supporting medium can effectively remove nitrate^{7,8,9,10}. The biological approach, which uses nitrate as a final electron acceptor in anoxic environments through microbial respiration, is thought to be the most practical and successful. Thus, biological treatment is more effective due to its potential to degrade almost all the pollutants while producing innocuous end products, reduce capital and operating costs, and maintain pollutant concentrations below the limit in the treated effluent. Denitrification can occur in the presence of oxygen in certain species too. Most of the organisms known to denitrify belong to bacterial genera belonging to *Pseudomonas*, *Rhizobium*, *Halobacterium*, *Halomona*, *Hyphomicrobium*, *Janthinobacterium*, *Neisseria*, *Paracoccus* (formerly *Micrococcus*), *Achromobacter*, *Acinetobacter*, *Aeromonas*, *Gallionella*, *Propionibacterium*, *Rhodobacter* (formerly *Rhodopseudomonas*), *Thiobacillus*, *Thiosphaera*, *Vibrio*, and *Xanthomonas*^{11, 12, 13}.

The reactors as MBR, SBR, packed bed reactors, and pulse plate reactors used for treating wastewater include non-ideality (dead/stagnant zones, channeling), uneven solid-liquid-gaseous phase mixing, increased energy needs, and higher fixed and recurrent costs¹⁴. The rate of oxygen and mass transport is subsequently impacted by these restrictions. Fluidized bed reactors for denitrification have been extensively studied by various researchers^{15, 16, 17}, and have shown some limitations like maximum bed expansion and retention time. The more nitrate is utilized at a lowest aeration frequency, with anoxic conditions in a draft tube being achieved in the annular part of the reactor¹⁸.

The aerobic biological treatment plant of industrial wastewaters is effectively being used with gas-liquid-solid spouted bed bioreactor¹⁹. The advantages of the spouted bed bioreactor are higher interfacial velocities, good mass transfer, and a higher biofilm liquid interfacial area. The

extreme increase of biomass on the media leads the washout of bioparticles from a reactor²⁰. The high flow rates in the spouted bed with the least jetting velocity lower the bed pressure drop and hence reduce the cost of operation²¹. Thus, a spouted bed reactor has been tested for its performance in the denitrification process in the present study using immobilized cells of *Pseudomonas syringae* as an isolate used for nitrate biodegradation.

Material and Methods

Microorganism

A strain of *Pseudomonas syringae* was stored at 4°C, which was isolated from the wastewater plant of a fertilizer industry. It was sub-cultured once in 15 days on a nutrient agar plate and used for further studies²².

Growth media composition

The growth media composition, which included nitrate-rich media and trace element solution, was listed in Table 1^{23, 24} along with the composition considered for 1 liter of water. The growth medium prepared was used in the experimental studies.

Table 1: Growth media composition

Nutrient rich media	Composition
NH ₄ Cl	0.3 gm/L
KH ₂ PO ₄	1.5 gm/L
Na ₂ HPO ₄ .7H ₂ O	7.9 gm/L
KNO ₃	2 gm/L
C ₄ H ₄ O ₄ .Na ₂ .6H ₂ O	27 gm/L
MgSO ₄ .7H ₂ O	20 gm/L
Trace element solution	Composition
EDTA	50.0 gm/L
ZnSO ₄	2.2 gm/L
CaCl ₂	5.5 gm/L
MnCl ₂ .4H ₂ O	5.06 gm/L
FeSO ₄ .7H ₂ O	5.0 gm/L
(NH ₄) ₆ Mo ₇ O ₂₄ .4H ₂ O	1.1 gm/L
CuSO ₄ .5H ₂ O	1.57 gm/L
CoCl ₂ .6H ₂ O	1.61 gm/L

Experimental bioreactor

The detailed representation of the draft tube-spouted bed reactor was as shown in Figure 1. The medium, all connecting tubes and other accessories were sterilized. Synthetic and growth media were loaded in the reactor along with immobilized granular activated carbon with *P. syringae*. To

have the controlled rate of proper supply of oxygen to a microorganism with spouting of solid, the compressed air was introduced through the filter.

The occupied volume of the reactor was 1.2 liters with draft tube length 45 cm and diameter 1.2 cm.

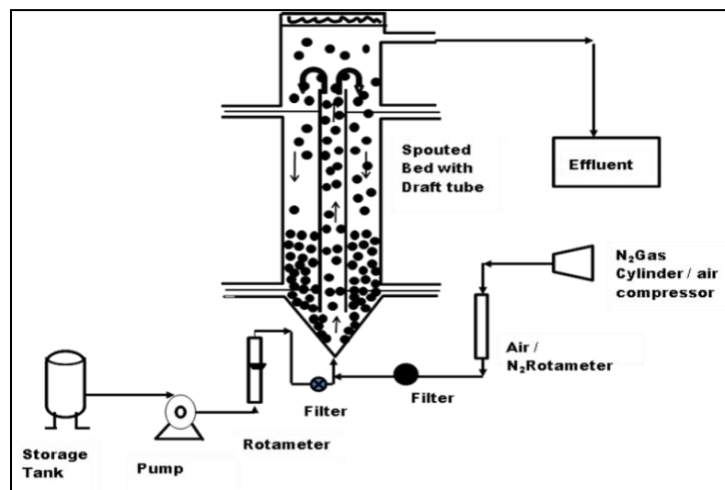


Figure 1: Schematic diagram of experimental setup.

Analytical methods

The sampling was done from the draft tube reactor for every hour to study the characteristics of the sample and hence the performance of the reactor. The sample was centrifuged (REMI, India) at a speed of 10000 rpm for a duration of 10 min, and the supernatant obtained was used for the analysis of nitrate and nitrite. A UV spectrophotometer was used to analyse both nitrate and nitrite. The solids were washed with 0.25 N NaOH solution and were dried at 105 °C for 24 h for analysing the attached biomass. The scanning electron microscope (SEM) was used to measure the biofilm thickness²².

Granular activated load (GAC)

GAC was procured from NICE Chemicals, Cochin, India. The GAC are screened for the desired size. The particles retaining on a 2 mm screen (average size = 2.4 mm) were taken for the study. The GAC was washed repeatedly using hot distilled water to remove all dirt, dust, and loose particles. Then, it was dried in an oven for two days at 105°C. This dried GAC was then used for

immobilization. The properties of GAC used in the study are given in Table 2.

Table 2: Properties of granular activated carbon used in the study

Sl. No	Properties	Value
1	Average size (mm)	2.4
2	Bulk density (kg/m ³)	610
3	True density (kg/m ³)	1278
4	Specific surface area (m ² /g)	680
5	Average pore radius (µm)	12
6	Pore volume (cm ³ /g)	0.45

Results

Study of nitrate removal percentage during start-up of the reactor with varying dilution rate.

It is evident from Figure 2 that, given an inlet nitrate concentration feed of 300 mg/L, the reactor's effluent nitrate concentration decreased continuously during the first phase of operation before slowing down for all dilution rates. The steady state was reached when the dilution rate was 0.166/h at 40th h, 0.333/h at 45th h, and 0.5/h at 50th h.

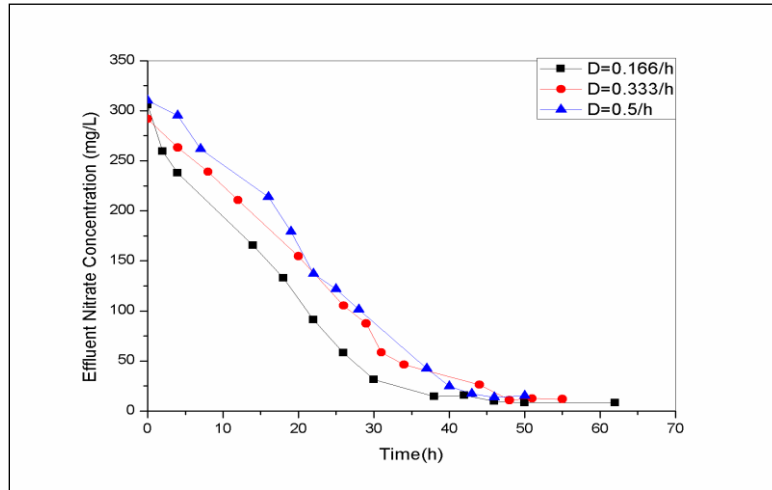


Figure 2: Study of nitrate removal percentage during start-up of the reactor with varying dilution rate with GAC loading of 40 g and nitrate concentration of 300 mg/L.

Study of attached biomass on percent nitrate removal for different dilution rate

It is evident from Figure 3 that the percentage of nitrate removal decreased as the amount of connected biomass increased. When the attached biomass was increased to 0.54 g/g with a dilution

rate of 0.166/h, the percentage of nitrate removal dropped to 88.2% from 98.18% at 0.35 g/g (Figure 3). The nitrate removal was 99% with a dilution rate of 0.166/h and an attached biomass of 0.35 g/g GAC, 94% for a dilution rate of 0.5/h, and the same attached biomass quantity.

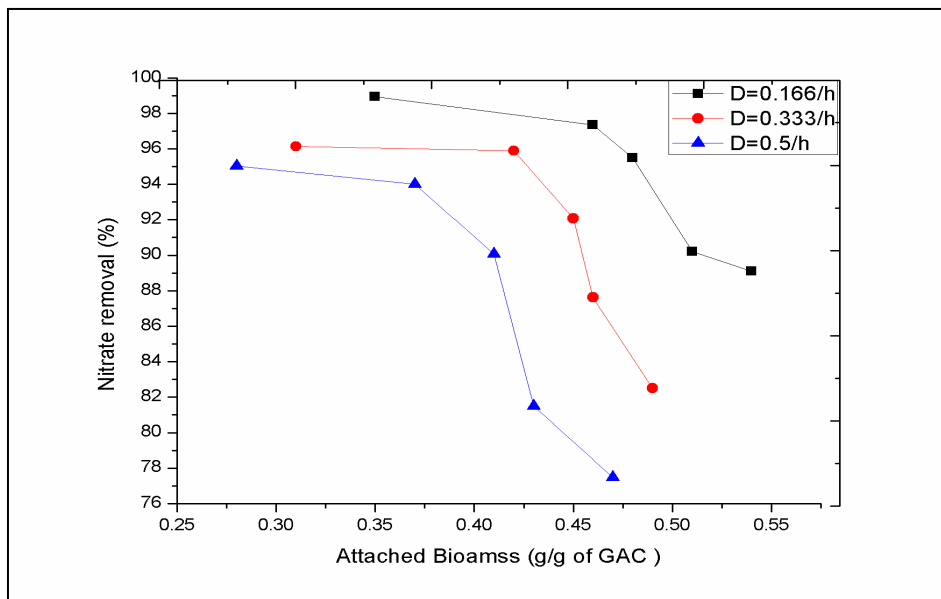


Figure 3: Study of attached biomass on percent nitrate removal at 40 g/g GAC loading.

Study of attached biomass on dilution rate and influent nitrate concentration.

It is observed from Figure 4 that the attached biomass quantity reduced as the dilution rate was increased with the initial concentration being

lowered. The attached biomass weight was 0.35 g for the initial nitrate concentration of 100 mg/L and dilution rate of 0.166/h. When the dilution rate was increased to 0.5/h, the attached biomass weight dropped to 0.27 g for the same initial nitrate

concentration. As the initial nitrate concentration rises, so does the attached biomass weight. The attached biomass weight was 0.35 g for the 100 mg/L initial nitrate concentration at the dilution

rate of 0.166/h, and it rose to 0.55 g for the starting nitrate concentration of 1200 mg/L at the same dilution rate.

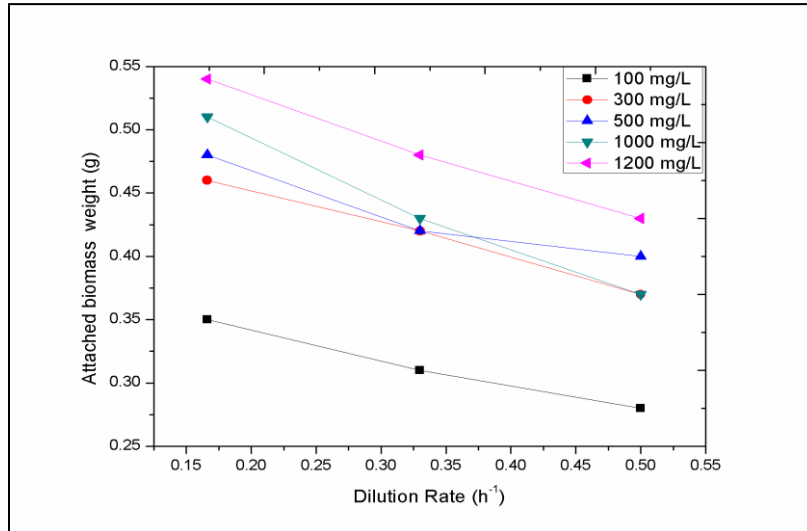


Figure 4: Study of attached biomass on dilution rate and influent nitrate concentration.

Study of biofilm thickness for different dilution rate and influent nitrate concentration.

It is observed from Figure 5 that the thickness of the biofilm rose as the concentration of influent nitrate increased and dropped as the rate of dilution

increased. It is evident that the biofilm thickness was 0.19 μm for an 100 mg/L initial nitrate concentration at a dilution rate of 0.166/h and 5.57 μm for an 1200 mg/L initial nitrate concentration at the same dilution rate.

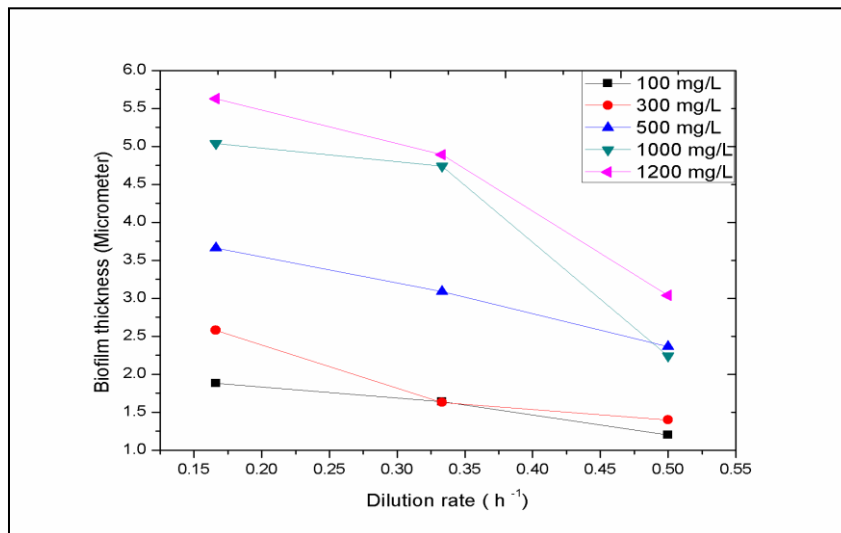


Figure 5: Study of biofilm thickness for different dilution rate and influent nitrate concentration.

Study of suspended biomass for different dilution rate and initial nitrate concentration.

From Figure 6, it is evident that when the

dilution rate and influent nitrate content increases, so does the suspended biomass. As can be seen, the suspended biomass was 4 g at 100 mg/l and rose to

10.5 g at 1200 mg/L of nitrate at a dilution rate of 0.166/h.

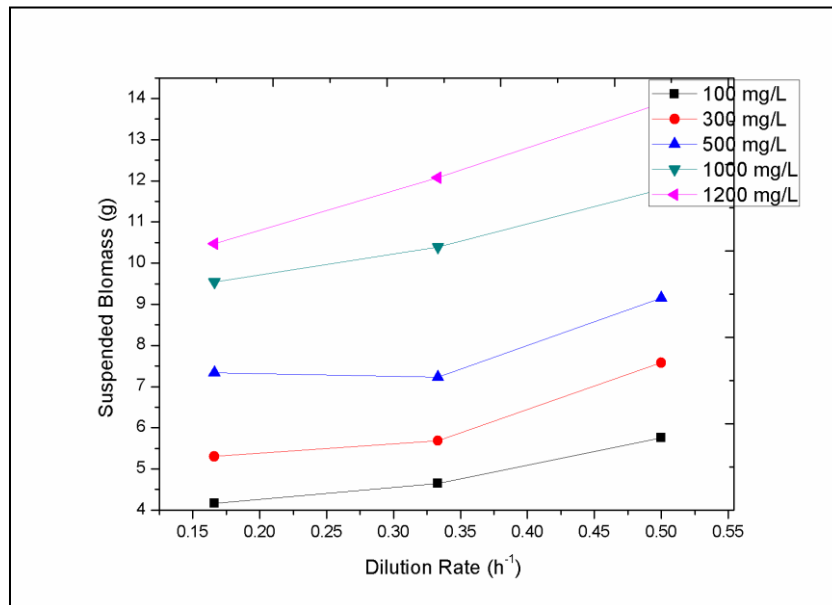


Figure 6: Study of suspended biomass for different dilution rate and initial nitrate concentration.

Morphological characteristics of biofilm and EPS formation

When examining the impact of biofilm thickness on biomass at varying dilution rates, it was observed that biofilm thickness decreased as the

dilution rate rose. According to Figure. 7, the biofilm thickness was 7.02 μm, 6.76 μm, and 5.56 μm for dilution rates of 0.166/h, 0.333/h, and 0.5/h, respectively.

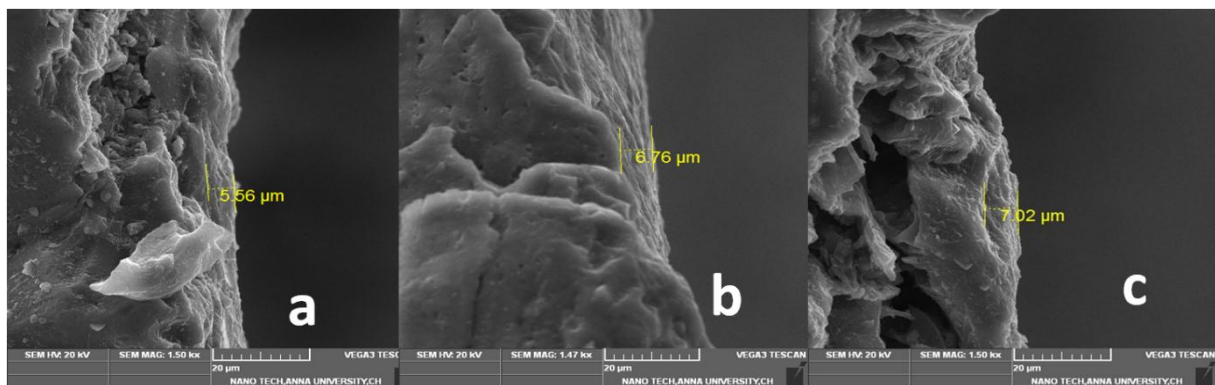


Figure 7: Morphological characteristics of biofilm (a) 0.5/h (b) 0.333/h (c) 0.166/h

Discussions

The study of dilution rate on nitrate reduction:

Nitrate adsorption onto the biofilm surface could be the cause of the initial abrupt decline. Cell walls, cell membranes, cytoplasm, and extracellular polymeric materials are potential sorption sites in biofilms. According to Aksu, these

sites support the sorption capabilities of biofilms for both organic and inorganic substances²⁵. It has been found that the organic compounds and microbial cell walls interact physiochemically to speed up the sorption step²⁶. The amount of nitrate removed by biodegradation may be less substantial during this stage. These cells are subjected to an

increase in shear stress, flowing liquid, and continually flowing air when they are placed in a continuous reactor after being acclimated, grown, and immobilized in a batch setting. The abrupt rise of shear stress on a biofilm stimulates the secretion of EPS-polysaccharides, resulting in sloughing of the biomass and attached biomass decreases in the adsorption phase^{27, 28}.

The study of attached biomass on nitrate reduction:

Since nitrate is one of the nutrients that increases metabolic activity, an increase in the concentration of nitrate in the inlet results in more nitrate available for the microorganisms and, consequently, higher growth. On GAC particles, this increased biomass development creates a denser and thicker biofilm. A lower percentage of nitrate is removed because of the thicker and denser biofilm's increased barrier to mass transfer²⁹. The flow velocity past the particle increases with the dilution rate. It is possible that the higher flow velocity caused biomass to be eroded off the film surface³⁰. The reduction in attached biomass with increased dilution rate is due to increased cell detachment by erosion.

The study of attached biomass on dilution rate on nitrate reduction:

Due to attrition between GAC particles and biofilm shearing, the amount of connected biomass reduced as the dilution rate increased^{31, 32}. Additionally, it is evident that a higher influent nitrate content resulted in more attached biomass since there were more nutrients available for biofilm formation. The stream velocity near the particle increases as the dilution rate rises. Higher biomass degradation from the biofilm surface may have been caused by this increased flow velocity³³.

The study of biofilm thickness on dilution rate:

The continuous flow of GAC particles in the draft tube part of the spouted bed reactor causes attrition and shear, which in turn results in the sloughing of associated biomass from the biofilm. The conditions under which the biofilm is produced determine its structure and morphology. The cohesion forces are provided by the EPS molecules

on the biofilm and adhesion to the substratum, and hence the stress distresses the composition^{34, 35, 36}. The detachment of the biomass occurs when the tensile strength of the EPS matrix is affected by tensile forces caused by the external shear^{37, 38}.

Effect of dilution rate on suspended biomass: More nutrients become available as the initial nitrate concentration rises, which promotes the growth of biomass³⁹. A higher dilution rate will result in more particle attrition, which will shear the biomass from the solids more and cause more biomass to wash out of the reactor⁴⁰.

Effect of Morphology and EPS Formation: The dry weight of the attached biofilm rose as the bacteria multiplied during the first phase, progressively increasing the thickness over time. The biomass content was directly correlated with the thickness of the biofilm; as biofilming grew, dry density dropped, increasing the biofilm's porosity and resulting in increased roughness⁴¹. It was found that when the dilution rate decreased, the nitrate reduction rate rose. Biofilm thickness rises with decreasing dilution rate, which enhances EPS generation and, consequently, nitrate reduction^{42, 43}.

The EPS has a significant impact on the structure, adsorption capacity, dehydration, and other chemical and physical characteristics of the biofilm. Because there was less cell decomposition in the current study, there was less organism degradation, which resulted in the production of humic material. It was observed that the production of EPS rose in tandem with the rate of nitrate reduction over time. Moreover, the nitrate that the microbes used to enable cell proliferation resulted in the formation of EPS in the first place, overlaying the biofilm and producing a significant quantity of EPS. As a result, the quantity of EPS rose over time with each component. After 70–80 hours, it was found that the EPS stayed consistent, signifying that the steady state has been achieved by the bioreactor has reached a steady state. A higher dilution rate would cause the biofilm to shred, which would minimize the generation of EPS. Additionally, a shorter residence time would result in less nitrate-cell interaction. The studies

have found that EPS produced largely polysaccharides in pure culture⁴⁴ and proteins in the mixed culture system⁴⁵.

Conclusion

The bioreactor chosen in the present study is complex in terms of circulation, mixing, holdup (hydrodynamics), DO level variation in the draft tube and annular region, and mass transfer aspects in terms of resistances in the bulk on the GAC surface and intraparticle diffusional resistance. The investigation shows that the rate of bio-denitrification reduces during start-up as the dilution rate increases. The time to attain a steady state is enhanced by an increase in dilution rate and influent nitrate concentration. At 300 mg/L influent nitrate concentrations and a dilution rate of 0.166/h, the bio-denitrification achieved was 98%. The biofilm thickness and attached biomass dry weight reduce with an increase in dilution rates as reduced cell growth and erosion are caused by increased cell detachment. The increase in dilution rate from 300 to 1200 mg/L increases the biofilm thickness and biomass weight due to the rise in growth rate of organisms at higher nitrate concentrations. The EPS formation has a significant impact on the bio-denitrification, resulting in minimizing the EPS at higher dilution rates. Also, the biofilm thickness rises with decreasing dilution rate, which enhances EPS generation and, consequently, nitrate reduction. Hence, a draft tube-spouted bed reactor could be a promising and sustainable approach to nitrate reduction.

Acknowledgement

The authors wish to thank the Department of Chemical Engineering, Management, and Principal S.D.M College of Engineering and Technology Dharwad for their support and encouragement.

Authors' contributions

All authors have contributed to the study conception and design. Material preparation, data collection and analysis were performed by Dr. Keshava Joshi. The first draft of the manuscript was written by Dr. Lokeshwari Navalgund and Dr Vinayak Shet, Dr. Keshava Joshi commented on

previous versions of the manuscript. All authors read and approved the final manuscript.

Authors ethically approve the submission and declared that it was not submitted elsewhere.

Competing interests

The authors declared no conflict of interests.

This is an Open-Access article distributed in accordance with the terms of the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt, and build upon this work for commercial use.

References

1. Arabzadeh M, Eslamidoost Z, Rajabi S, et al. WasteWater Quality Index (WWQI) as an indicator for the assessment of sanitary effluents from the oil and gas industries for reliable and sustainable water reuse. *Groundw Sustain Dev.* 2023; 23: 101015.
2. Karanasios K, Vasiliadou I, Pavlou S, et al. Hydrogenotrophic denitrification of potable water: a review. *J Hazard Mater.* 2010; 180: 20–37.
3. Hashemi H, Rajabi S, Nikooee S, et al. Ozonation of secondary industrial effluent for beneficial reuse. *Desalination Water Treat.* 2023; 287: 96–102.
4. Shrimali M, Singh K. New methods of nitrate removal from water. *Environmental Pollution.* 2001; 112(3): 351–9.
5. Foglar L, Briski F, Sipos L, et al., High nitrate removal from synthetic wastewater with the mixed bacterial culture. *Bioresour Technol.* 2005; 96(8): 879–88.
6. Chibata I, Wingard LB. Immobilized microbial cells, Applied Biochemistry and Bioengineering. New York: Academic Press. 1983;4.
7. Santos VA, Tramper J, Wiffels RH. Simultaneous nitrification and denitrification using immobilized microorganisms. *Biomater Artif Cells Immobilization Biotechnol.* 1993; 21(3): 317–22.
8. Uemoto H, Saiki H. Distribution of *Nitrosomonas europaea* and *Paracoccus denitrificans* immobilized in tubular polymeric gel for nitrogen removal. *Appl Environ Microbiol.* 2000; 66(2):

- 816-9.
9. Kesseri P, Kiss I, Bihar Z, et al., Investigation of the denitrification activity of immobilized *Pseudomonas butanovora* cells in the presence of different organic substrates. *Water Res.* 2002; 36(6): 1565-71.
 10. Kube M, Spedding B, Gao L, et al. Nutrient removal by alginate-immobilized *Chlorella vulgaris*: response to different wastewater matrices. *Journal of Chemical Technology & Biotechnology.* 2020; 95(6): 1790-9.
 11. Mateju V, Cizinska S, Krejci J, et al. Biological water denitrification – a review. *Enzyme Microb Technol.* 1992; 14(3): 170-83.
 12. Mansell BO. Biological denitrification of nitrate contaminated groundwater in a microporous membrane bioreactor. University of California, Davis. 2000.
 13. Fendrihan S, Negoita TG. Psychrophilic microorganisms as important source for biotechnological processes. *Adaption of Microbial Life to Environmental Extremes: Novel Research Results and Application.* 2017; 147-99.
 14. Kavitha E, Poonguzhali E, Nanditha D. et al. Current status and future prospects of membrane separation processes for value recovery from wastewater. *Chemosphere.* 2022; 291: 132690.
 15. Rabah FK, Dahab MF. Nitrate removal characteristics of high performance fluidized-bed reactors. *Water Res.* 2004; 38 (17): 3719-28.
 16. Toumi LB, Fedailaine M, Khedidja A. Modeling three phase fluidized bed bioreactor for wastewater treatment. *International Journal of Chemical Reactor Engineering.* 2008; 6(1): 1542-6580.
 17. Prasanna L, Pydi Setty Y, Vimala K. A Study on Denitrification in a Fluidized bed Bioreactor, ECI Conference on the 13th International Conference on Fluidization - New Paradigm in Fluidization Engineering. 2010.
 18. Dhamole PB, Nair RR, D'Souza SF, et al. Denitrification of high strength nitrate waste. *Bioresour Technol.* 2007; 98(2): 247-52.
 19. Dabhade MA, Saidutta MB, Murthy DV. Continuous phenol removal using *Nocardia hydrocarbonoxydans* in spouted bed contactor: Shock load study. *Afr J Biotechnol.* 2009; 8 (4): 644-9.
 20. Sokol W. Operational range for a gas-liquid-solid fluidized bed aerobic biofilm reactor with a low-density biomass support. *International Journal of Chemical Reactor Engineering.* 2010;8(1).
 21. Vukovic DV, Zdanski FK, Vunjak GV, et al. Pressure drop, bed expansion and liquid hold up in a three-phase spouted bed contactor. *Can J Chem Eng.* 1974; 52(2): 180-4.
 22. Joshi K, Joseph J, Srinikethan G, et al. Isolation and characterization of *Pseudomonas syringae* for nitrate removal under aerobic conditions. *J Biochem Technol.* 2014; 5(2):693-7.
 23. Su JJ, Liu BY, Liu CY. Comparison of aerobic denitrification under high oxygen atmosphere by *Thiosphaera pantotropha* ATCC 35512 and *Pseudomonas stutzeri* SU2 newly isolated from the activated sludge of a piggery wastewater treatment system. *J Appl Microbiol.* 2001; 90(3): 457-62.
 24. Kim M, Jeong SY, Yoon SJ. et al. Aerobic denitrification of *Pseudomonas putida* AD-21 at different C/N ratios. *J Biosci Bioeng.* 2008; 106(5): 498-502.
 25. Aksu Z. Application of biosorption for the removal of organic pollutants: a review. *Process Biochemistry.* 2005; 40(3-4): 997–1026.
 26. Mark CM, Van L, Johannes L, et al. Influence of Interfaces on Microbial Activity. *Microbiol Rev.* 1990; 54(1): 75-87.
 27. Nicolella C, Chiarle S, Felice RD, et al. Mechanism of biofilm detachment in fluidized bed reactors. *Water Sci Technol.* 1997; 36 (1): 229–35.
 28. Ramasamy P, Zhang X. Effects of shear stress on the secretion of extracellular polymeric substances in biofilms. *Water Sci Technol.* 2005; 52 (7): 217-23.
 29. Gibert O, Lefevre B, Fernandez M, et al. Characterizing biofilm development on granular activated carbon used for drinking water production. *Water Res.* 2013; 47(3): 1101-10.
 30. Vidya shetty K, Ramanjaneyulu R, Srinikethan G. Biological phenol removal using immobilized

- cells in a pulse palte bioreactor: effect of dilution rate and influent phenol concentration. *J Hazard Mater.* 2007; 149(2): 452-9.
31. Joshi K, Navalgund L, Rathod K, et al. Biofilm characterization in removal of total chemical oxygen demand and nitrate from wastewater using draft tube spouted bed reactor. *Biotechnol Lett.* 2021; 43: 2001-9.
 32. Turan M. Backwashing of granular media filters and membranes for water treatment: a review. *AQUA—Water Infrastructure. Ecosystems and Society.* 2023; 72(3): 274-98.
 33. Picioreanu C, Mark CM, Loosdrecht V, et al. Effect of diffusive and convective substrate transport on biofilm structure formation: a two-dimensional modeling study. *Biotechnology and Bioengineering.* 2000; 69(5): 504-15.
 34. Salama Y, Chennaoui M, Sylla A, et al. Characterization, structure, and function of Extracellular Polymeric Substances (EPS) of microbial biofilm in biological wastewater treatment systems: a review. *Desalination Water Treat.* 2016; 57(35): 16220-37.
 35. Melo A, Quintelas C, Ferreira EC, et al. The role of extracellular polymeric substances in micropollutant removal. *Front Chem Eng.* 2022; 4: 778469.
 36. Schmitt J, Nivens D, White DC, et al. Changes of biofilm properties in response to sorbed substances - an Am-ATR study. *Water Sci Technol.* 1995; 32(8):149-55.
 37. Ohashi A, Harada H. Adhesion strength of biofilm developed in an attached-growth reactor. *Water Sci Technol.* 1994; 29 (10): 281-8.
 38. Kwok WK, Picioreanu C, Ong SL, et al. Influence of biomass production and detachment forces on biofilm structures in a biofilm airlift suspension reactor. *Biotechnology and Bioengineering.* 1998; 58(4): 400-7.
 39. Ahmed SF, Mofijur M, Parisa TA. et al. Progress and challenges of contaminate removal from wastewater using microalgae biomass. *Chemosphere.* 2022; 286: 131656.
 40. Wlodzimierz S. Optimal aerations in the inverse fluidized bed biofilm reactor when used in treatment of industrial wastewaters of various strength. *Advances in Chemical Engineering and Science.* 2012; 2(3): 384-91.
 41. He J, Gao X, Huang H, et al. Proposal and verification of the theory of layer-by-layer elimination of biofilm in listeria monocytogenes. *Foods.* 2023; 12(7): 1361.
 42. Cao S, Peng Y, Du R, et al. Characterization of Partial-Denitrification (PD) granular sludge producing nitrite: effect of loading rates and particle size. *Sci Total Environ.* 2019; 671: 510-8.
 43. Wu D, Zhao B, Zhang P, et al. Insight into the effect of nitrate on AGS granulation: granular characteristics, microbial community and metabolomics response. *Water Res.* 2023; 236: 119949.
 44. Veena BR, Shetty KV, Saidutta MB. Shear stress effects on production of exopolymeric substances and biofilm characteristics during phenol biodegradation by immobilized *Pseudomonas desmolyticum* (NCIM2112) cells in a pulsed plate bioreactor. *Prep Biochem Biotechnol.* 2016; 46(5): 421-8.
 45. Salama Y, Chennaoui M, Sylla A, et al. Characterization, structure, and function of Extracellular Polymeric Substances (EPS) of microbial biofilm in biological wastewater treatment systems: a review. *Desalination Water Treat.* 2016; 57(35): 16220-37.