

Evaluation of Oxidative Stress Induced by Occupational Inhalation Exposure to N₂O, an Anesthetic Gas

Masoud Neghab¹, Fatemeh Kargar-Shouroki^{2*}, Saeed Yousefinejad³, Hamzeh Alipour¹, Hossein Mozdarani⁴, Reza Fardid⁵, Vida Sadat Anoosheh^{6,3}, Masoud Rostami⁷

¹ Research Center for Health Sciences, School of Health, Shiraz University of Medical Sciences, Shiraz, Iran.

² Industrial Diseases Research Center, Department of Occupational Health Engineering, School of Public Health, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

³ Department of Occupational Health Engineering, School of Health, Shiraz University of Medical Sciences, Shiraz, Iran.

⁴ Department of Medical Genetics, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran.

⁵ Department of Radiology, School of Paramedical Sciences, Shiraz University of Medical Sciences, Shiraz, Iran.

⁶ Student Research Committee, Shiraz University of Medical Sciences, Shiraz, Iran.

⁷ Department of Languages and Literature, Yazd University, Yazd, Iran.

ARTICLE INFO

ORIGINAL ARTICLE

Article History:

Received: 17 November 2023

Accepted: 20 January 2024

*Corresponding Author:

Fatemeh Kargar-Shouroki

Email:

kargar_st@yahoo.com

Tel:

+98 35 38209100

Keywords:

Nitrous Oxide,
Operating Rooms,
Oxidative Stress,
Ventilation,
Anesthetics.

ABSTRACT

Introduction: Nitrous oxide (N₂O) is the most common anesthetic gas used in operating rooms. The major objective of this investigation is to measure N₂O values in two modes: first, when the ventilation system is on, and second, when it is off; and to determine the biomarkers of oxidative stress associated with this exposure among operating room personnel.

Materials and Methods: A cross-sectional study was conducted on 60 operating room personnel as the N₂O exposed group, and on 60 nurses as the referent group. N₂O concentrations were determined according to NIOSH method 6600. Total antioxidant capacity (TAC) levels, malondialdehyde (MDA), and superoxide dismutase (SOD) activities were also measured.

Results: The concentrations of N₂O in the presence and absence of ventilation systems were significantly higher than the recommended exposure limit (REL) of 25 ppm recommended by NIOSH. The levels of TAC and SOD were significantly lower in participants exposed to N₂O in comparison with the referent group. Adjusted for age, work experience, and sex, exposure to N₂O was found to be an occupational risk factor for low levels of TAC and SOD, so that exposure to N₂O reduced TAC and SOD levels by 0.16 mM and 0.75 U/ml, respectively.

Conclusion: The present study shows that the operating room personnel are exposed to levels of N₂O several times more than the REL of this gas and this heavy exposure is associated with a significant increase in oxidative stress.

Citation: Neghab M, Kargar-Shouroki F, Yousefinejad S, et al. *Evaluation of Oxidative Stress Induced by Occupational Inhalation Exposure to N₂O, an Anesthetic Gas*. J Environ Health Sustain Dev. 2024; 9(1): 2205-13.

Introduction

Nitrous oxide (N₂O) is a non-flammable and colorless gas which induces rapid anesthesia without skin or trachea irritation. However, there have been concerns about toxic effects of N₂O

among operating room personnel who are regularly exposed to different values of N₂O for several years since the mid-1950s¹.

Chronic exposure to anesthetic gases affect the developing of fetus,² and results in infertility³ in

operating room personnel. Using employment records, Teschke et al. conducted a study through a telephone survey on Canadian nurses with a history of exposure to anesthetic gases during 1990-2000. They identified 1,079 cases of congenital malformations and 80 stillbirths among 15,317 live births recorded from 9,433 mothers. In mothers exposed to N₂O, congenital malformations were 1.42 times more than others⁴.

N₂O also affects liver,⁵ kidney,⁶ nerves system,⁷ and DNA^{8,9}.

Some studies have also reported the generation of oxidative stress (OS) by inhalational anesthetics. Oxidative stress is caused by an imbalance between the generation of reactive oxygen species (ROS) such as hydroxyl radical (HO[•]), superoxide anion (O₂^{•-}), hydrogen peroxide (H₂O₂), and antioxidant defense^{10,11,12}. Many studies have shown that ROS is related to reproductive, neurological, cardiovascular, and cancer diseases¹³.

ROS can damage proteins and nucleic acids¹³⁻¹⁶, it also affects the lipids of the cell membrane (lipid peroxidation) and leads to the formation of malondialdehyde (MDA), which is the most significant measure of lipid peroxidation¹².

Some studies have shown that anesthetic gases increase MDA levels in the operating room personnel^{6,14}. Significant lower levels of superoxide dismutase (SOD) and higher MDA were reported in the personnel exposed to anesthetics in comparison with the non-exposed group¹⁷.

The N₂O indirectly induces genotoxicity through ROS mediators¹⁸. There is also some evidence that N₂O toxicity is related to vitamin B12 deficiency¹. Vitamin B12 helps DNA metabolism and methionine synthesis¹⁹. N₂O irreversibly oxidizes the cobalt atom in vitamin B12 (cobalamin) and inhibits methionine enzyme synthesis. Oxidized vitamin B12 by N₂O causes the creation of O₂^{•-} and HO[•] radicals that impairs the conversion of homocysteine to methionine. A high level of homocysteine decreases the expression of antioxidant-related genes,¹⁸ production of ROS, mitochondrial dysfunction, and DNA damage. These subtle changes are associated with observed adverse health effects such as genotoxicity,

neurotoxicity, and teratogenicity¹.

Enzymatic ROS scavenging like glutathione peroxidase (GPx), SOD, catalase, and non-enzymatic ROS scavenging such as vitamin E and vitamin C protect cells against mutations and reduce radical attack on DNA^{13,14,16,20}.

To prevent the adverse effects of N₂O, National Institute of Occupational Safety and Health (NIOSH) and American Conference of Governmental Industrial Hygienists (ACGIH) recommended a maximum permissible concentration of 25 ppm and 50 ppm nitrous oxide, respectively^{21,22}.

In Iran, most of general anesthesia is performed using N₂O²³. In this regard, the main objective of this study is to measure N₂O concentrations based on two modes: first, when the ventilation system is on, and second, when it is off and to measure the biomarkers of oxidative stress associated with this exposure among operating room personnel

Materials and methods

Study area and sampling

This study was conducted on 60 operating room personnel in Shiraz, Iran, and oxidative stress biomarkers were determined in these subjects. The inclusion criteria were at least 3 years of exposure to anesthetic gases and 36 to 44 hours of work per week during the past 3 months, except for weekends. All the operating room personnel worked 8 hours a day, except for surgeons who spent 6 hours a day in the operating room.

60 nurses without history of exposure to N₂O, who were compared to the exposed group in terms of age, sex, and work experience, were randomly selected from other departments of the same hospital as the referent group.

Ethics permit and questionnaire

Prior to conducting the research, ethical clearance was obtained from Ethics Committee of the Shiraz University of Medical Sciences. Demographic information and the data regarding work experience, alcohol consumption, smoking status, use of drugs including antibiotics, exposure to other chemicals causing oxidative stress, medical history, and detailed work history including job position and

working hours per day were collected through a questionnaire.

Participants who suffered from lung and liver diseases, chronic cardiovascular diseases, autoimmune, inflammatory, digestive, neurological diseases, diabetes, cancer, high blood pressure, had a BMI of more than 30 kg/m², people with a history of surgery, those with acute infections who needed to take drugs such as antibiotics during the last three months, those who used antioxidants such as vitamins E and C and the subjects with working hours of less than 6 hours a day were excluded from the study.

Exposure assessment

Exposure to N₂O was measured by NIOSH 6600 method²⁴. The concentration of N₂O gas was measured in 8 operating rooms and 1 recovery room under conditions where the ventilation systems were either on or off. Collectively 900 measurements in 8 operating rooms and 1 recovery room in morning shifts during a period of 2 months were conducted. To determine N₂O gas, an IR spectrophotometer (Bacharach model 3010, New Kensington, PA, USA) was used, which was field readout device. The sampling collection points were:

- From a distance of 15 cm from the breathing zone of the operating room personnel
- From a distance of 15 cm from the breathing zone of the recovery personnel
- From a distance of 5 cm from the tracheal tube or anesthesia mask of the patients
- From a distance of 5 cm from the anesthesia machine

- From a distance of 5 cm from the exhaust air grille
- From a distance of 15 cm from the breathing zone of the referent group

Evaluation of antioxidant status

Blood samples were collected from studied groups and transferred to tubes, and after clotting, they were centrifuged at 1200 rpm for 10 minutes so that the sera were separated. Serum specimens were kept in a refrigerator at -80°C until analysis.

MDA, SOD, and TAC were measured using kit (Zellbio Lab, Ulm, Deutschland, Germany) according to the manufacturer's protocol on a Stat FAX 2100 ELISA plate reader (Awareness Inc, USA).

Statistical analysis

Data was managed employing SPSS software, and chi-square test was used to assess the distribution of categorical variables. Independent t-test and one-way ANOVA were used to compare the quantitative indicators between the two and the more than two groups, respectively. A multivariate linear regression model was used to evaluate the association between oxidative stress status and N₂O exposure after adjustment for the effect of confounders (age, work experience, and sex).

Results

Table 1 shows some of the main characteristics of the studied groups. There were no significant differences regarding age, work experience, BMI, sex, and marital status between the groups ($p > 0.05$).

Table 1: Demographic information of the subjects

Variables	Referent group Mean ± SD	Exposed group Mean ± SD			P-value
		Technician	Nurses	Surgeons	
Age (year)	35.13 ± 6.36	37.81 ± 6.97	34.15 ± 6.76	40.40 ± 8.19	0.06*
Work experience (year)	10.27 ± 6.03	12.67 ± 5.39	9.98 ± 5.51	11.50 ± 5.93	0.42*
BMI (kg/m ²)	20.99 ± 3.14	20.56 ± 3.54	20.73 ± 2.45	21.10 ± 2.90	0.94*
Sex	Number (percent)				
Male	35 (58.30)	5 (31.20)	18 (52.90)	7 (70)	0.36**
Female	25 (41.70)	11 (68.80)	16 (47.10)	3 (30)	
Marital status	Number (percent)				
Single	13 (21.70)	2 (12.50)	7 (20.60)	1 (10)	0.73**
Married	47 (78.30)	14 (87.50)	27 (79.40)	9 (90)	

*One-way ANOVA

**Chi-square test

Table 2 shows the mean N₂O concentrations at 6 measured points at ppm range. The mean concentrations of N₂O in the off mode of ventilation were 582.33 ± 87.38, 263.06 ± 15.22, 1135.27 ± 48.31, 1905.57 ± 130.48, and 2219.98 ± 233.45, and not detectable for measurement points 1 to 6,

respectively. The corresponding values in the on mode of ventilation system were 241.49 ± 63.67, 118.50 ± 13.17, 707.43 ± 42.93, 1523.63 ± 125.67, and 1412.64 ± 102.39, and not detectable, respectively. These differences were statistically significant (p < 0.05).

Table 2: Mean concentrations of nitrous oxide (ppm) according to the ventilation system at different points

Measurement points	Number	Ventilation system off Mean ± SD	Number	Ventilation system on Mean ± SD	P-value*
1 From a distance of 15 cm from the breathing zone of the operating room personnel	200	582.33 ± 87.38	200	241.49 ± 63.67	< 0.001
2 From a distance of 15 cm from the breathing zone of the recovery personnel	50	263.06 ± 15.22	50	118.50 ± 13.17	< 0.001
3 From a distance of 5 cm from the anesthesia mask	50	1135.27 ± 48.31	50	707.43 ± 42.93	0.01
4 From a distance of 5 cm from the exhaust air grille	50	1905.57 ± 130.48	50	1523.63 ± 125.67	0.01
5 From a distance of 5 cm from the anesthesia machine	50	2219.98 ± 233.45	50	1412.64 ± 102.39	0.02
6 From a distance of 15 cm from the breathing zone of the referent group	50	ND	50	ND	-

* Independent sample t-test

ND: Not detectable

Oxidative stress biomarkers in the studied groups based on demographic data are summarized in Table 3. MDA and TAC levels and SOD activities did not change significantly with age and work experience. However, When SOD and TAC were separated by sex, males had lower SOD and

TAC levels than females in the exposed and referent groups, respectively. (SOD: 9.72 ± 3.73 U/ml vs. 12.72 ± 5.87 U/ml in the males and females exposed group, respectively) (TAC: 1.86 ± 0.50 mM vs. 2.32 ± 0.67 mM in the males and females of the referent group, respectively).

Table 3: Oxidative stress biomarkers of the studied groups based on demographic data

Variables	Referent group			n	Exposed group			
	MDA (μM)	SOD (U/ml)	TAC (mM)		MDA (μM)	SOD (U/ml)	TAC (mM)	
	Mean ± SD				Mean ± SD			
Age (year)								
< 35	28	2.19 ± 0.70	13.31 ± 3.72	2.10 ± 0.64	34	2.47 ± 0.68	11.14 ± 4.64	1.79 ± 0.64
≥ 35	32	2.32 ± 0.76	13.40 ± 4.49	2.16 ± 0.65	26	2.46 ± 0.65	11.33 ± 5.76	1.71 ± 0.54
		P = 0.50	P = 0.94	P = 0.76		P = 0.96	P = 0.89	P = 0.63
Work experience (year)								
≤ 10	34	2.28 ± 0.76	13.71 ± 4.43	2.12 ± 0.64	34	2.37 ± 0.67	12.15 ± 4.92	1.89 ± 0.63
> 10	26	2.24 ± 0.70	12.89 ± 3.70	2.15 ± 0.65	26	2.56 ± 0.66	10.23 ± 5.20	1.62 ± 0.52
		P = 0.87	P = 0.45	P = 0.88		P = 0.25	P = 0.15	P = 0.08
Sex								
Male	25	2.05 ± 0.81	13.26 ± 3.95	1.86 ± 0.50	34	2.46 ± 0.66	9.72 ± 3.73	1.75 ± 0.52
Female	35	2.41 ± 0.63	13.43 ± 4.28	2.32 ± 0.67	26	2.48 ± 0.67	12.72 ± 5.87	1.76 ± 0.67
		P = 0.07	P = 0.87	P = 0.004		P = 0.98	P = 0.02	P = 0.92

The average levels of TAC, SOD, and MDA based on job titles are given in Table 4. As shown, the lowest levels of TAC and SOD were observed

among the operating room nurses and technicians, respectively (1.70 ± 0.59 mM for TAC and 9.50 ± 3.46 U/ml for SOD).

Table 4: Oxidative stress biomarkers in the studied groups based on job titles

Groups	TAC (mM)	SOD (U/ml)	MDA (μM)
	Mean ± SD	Mean ± SD	Mean ± SD
Referent group	2.13 ± 0.64	13.36 ± 4.12	2.19 ± 0.68
Technicians	1.81 ± 0.60	$9.50 \pm 3.46^{***}$	2.32 ± 0.67
Operating room nurses	$1.70 \pm 0.59^{**}$	11.46 ± 5.30	2.58 ± 0.65
Surgeons	1.88 ± 0.63	13.18 ± 6.18	2.30 ± 0.67
P-value*	0.001	0.01	0.03

* One-way ANOVA

**A statistically significant difference was observed between operating room nurses and the referent group ($p = 0.02$).

***A statistically significant difference was observed between technicians and the referent group ($p = 0.03$).

Variables such as age, work experience, and sex were considered as confounders, and their effects on the oxidative stress were controlled using linear regression analysis. The model was made based on the main exposure variable, N₂O exposure, as well as all the confounding variables.

Table 5 presents the results obtained from the linear regression analysis. As shown in Table 5, there was a negative association between N₂O exposure and TAC and SOD levels. Exposure to N₂O resulted in a 0.16 mM and 0.75 U/ml decrease in TAC level and SOD activity, respectively (Table 5).

Table 5: Association between oxidative stress status and nitrous oxide exposure in the studied groups

Variables	B	95% confidence interval	P-value
TAC	-0.16	-0.27 to -0.06	0.002
SOD	-0.75	-1.53 to 0.03	0.05
MDA	0.80	-0.03 to 0.20	0.16

Discussion

N₂O gas is one of the most common pollutants in operating rooms. N₂O concentrations, in both on and off modes of ventilation system were several-fold higher than the REL value of 25 ppm suggested by NIOSH²¹ and the threshold limit value (TLV) of 50 ppm suggested by ACGIH²².

Similarly, Souza et al. and Braz et al. reported mean N₂O concentration of 170 ppm and 155 ppm in operating room personnel, respectively^{25, 26}.

Wiesner et al. in a study on hospitals with and without a ventilation system reported N₂O concentrations of 170 ppm and 12 ppm, respectively²⁷.

In the present study, N₂O concentrations were substantially higher in comparison with similar studies. The reasons could be improper ventilation and scavenging systems, gas leakage from the anesthesia machine and patient's mask, and large number and duration of daily surgeries²⁸.

In this study, the scavenging system to remove waste anesthetic gases and exhausts were improperly designed. Moreover, air circulations were less than the standard of 15 and 6 air changes per hour in operating and recovery rooms, respectively.

In this study, higher levels of TAC and SOD were observed in females compared with males. Similar findings had been reported by others^{29, 30}.

The followings are some of the reasons for the higher levels of TAC and SOD in women:

1- Female's mitochondria levels generate almost half the amount of hydrogen peroxide as compared to males.

2- Mitochondrial glutathione levels are almost twice in females than males.

3- Females overexpress SOD and glutathione peroxidase and mitochondrial enzymes²⁹, and this is due to estrogens that bind to estrogen receptors and activate the mitogen-activated protein (MAP) kinase and nuclear factor kappa (NF-κB) signaling pathways³¹.

4- Males also produce more ROS due to NADPH oxidase activity, which is a major oxidative stress producer in cells^{29, 30}.

5- Males have more homocysteine than females

due to their sex hormone (progesterone). Homocysteine is an indicator of folate and B-12 deficiency,³⁰ which also increases as a result of exposure to N₂O, and high levels of homocysteine lead to a decrease in the expression of antioxidant-related genes, production of ROS, mitochondrial dysfunction, and DNA damage¹⁸.

In the present study, operating room personnel had a lower mean regarding TAC and SOD levels in comparison with the referent group. This finding was in line with the finding of Turkan et al.³².

In 2005, Malekirad et al. reported a significant lower thiol groups and higher lipid peroxidation in 66 exposed operating room personnel with 9 years of work experience¹⁴.

Izdes et al. showed that exposure to anesthetic gases significantly reduced TAC and glutathione in comparison to the control group²⁰. Similar findings were reported in 2014 by Cerit et al.³³

Cegin et al. in a study on 32 operating room staff and 32 control groups in 2016, showed a significant increase in lipid peroxidation and serum myeloperoxidase activity as well as a decrease in catalase activity and sulfhydryl levels in the exposed group compared to the non-exposed group³⁴.

Similarly, according to Paes's study in 2014 on 15 medical residents in Brazilian hospitals¹⁰ and Baysal's research on 30 operating room personnel in Turkey, antioxidant capacities in operating room personnel were significantly lower than the control group³⁵.

In the study by Jafari et al. in Iran in 2018, an increase in MDA was observed following exposure to high levels of inhalational anesthetics⁶.

As oxidative stress induced by inhalational anesthetics is one of the mechanisms of DNA damage, the protective impacts of antioxidants had been confirmed in some studies on operating room personnel^{20, 36-38}. For example, Sardas et al. reported that the use of vitamins E and C by operating room personnel could reduce DNA damage in operating room staff in comparison with the control group³⁷. Similarly, a worldwide cohort study from 1989 to 2009, in which 16 countries and 5,424 people were studied, reported that the micronucleus frequency in subjects who consumed

fruits and vegetables once a day was 32% lower than those who reported no consumption³⁶. Similar results were reported in 2010 by Izdes et al. regarding operating room staff²⁰.

Ranjbar et al. reported that the daily consumption of 0.10 g of cinnamon in 100 cc of water for 10 days caused a significant decrease in the level of lipid peroxidation (3.25 ± 1.32 versus 5.03 ± 2.01 nmol/ml)³⁹.

Similarly, Sami et al. reported that daily consumption of two cups of tea (including 1.873 grams of chamomile in 300 cc of water) for 21 days significantly increased salivary TAC levels (6.62 ± 0.77 μ mol/ml versus 4.81 ± 0.39 μ mol/ml)⁴⁰.

Due to the inherent limitations of cross-sectional studies such as this one, it is impossible to assess a causal association. Thus, it may be argued that a significant reduction in antioxidant defense in operating room staff is not related to exposure to N₂O. Though from the point of view of epidemiology, this is correct, the authors believe that there are multiple proofs to prove that the observed effects may well be attributed to exposure to N₂O. These include:

- 1- There were no significant differences between demographic data of the studied groups.
- 2- The exposed group had no history of autoimmune disorders, inflammatory diseases, or lung or liver disorders that could affect oxidative stress.
- 3- The exposed group had no history of exposure to other physical and chemical factors causing oxidative stress biomarkers.
- 4- All the exposed groups were non-smokers.
- 5- Antioxidant biomarkers were significantly lower, while the oxidative stress biomarkers were significantly higher in the exposed group than in the referent group.
- 6- After adjusting for the confounders, the association between low levels of SOD and TAC and exposure to N₂O remained significant, indicating that the observed changes were associated with exposure to N₂O.

"A limitation of this study was that the effect of shiftwork was not studied. Therefore, more studies regarding the role of shiftwork are needed".

Conclusion

The present study shows a decrease in antioxidant status in operating room staff compared to the control group. Since there were no significant differences in the main variables of age, BMI, sex, smoking status, and work experiences between the studied groups, the observed effects can be associated to exposure to high levels of N₂O. Therefore, engineering and administrative control measures are suggested to reduce workers exposure to N₂O.

Acknowledgements

The authors would like to thank all the operating room personnel who participated in the study.

Conflict of interest

The authors declared no conflict of interest.

Funding

This work was supported by the Shiraz University of Medical Science (SUMS) under Grant (95-01-04-12366).

Ethical considerations

The study was conducted in accordance with the Helsinki Declaration of 1964 as revised in 2013.

Code of ethic

The protocol of the study was approved by Shiraz University of Medical Sciences ethics committee (IR.SUMS.REC.1395.S729).

Authors' contributions

Neghab M developed and designed the study, Kargar-Shouroki F collected and analyzed data, Yousefinejad S, Alipour H, Mozdarani H, Fardid R, Anoosheh VS wrote the first draft of the manuscript, and Rostami M revised the manuscript. All the authors read and approved the final manuscript.

This is an Open-Access article distributed in accordance with the terms of the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt, and build upon this work for commercial use.

References

1. Menon JM, van Luijk JA, Swinkels J, et al. A

- health-based recommended occupational exposure limit for nitrous oxide using experimental animal data based on a systematic review and dose-response analysis. *Environmental research*. 2021;201:1-16.
2. Ge B, Sa AE-M, Ss E-G, et al. occupational genotoxic effects among a group of nurses exposed to anesthetic gases in operating rooms at Zagazig university hospitals. *Egypt J Occup Med*. 2018;42(1):105-22.
 3. Handayani R, Abdullah T, Naiem F, et al. Effects of Isoflurane Exposure to Fertility through Estrogen Gene Expression in Operating Room Nurses. *Am J Public Health*. 2018;6(1):11-7.
 4. Teschke K, Abanto Z, Arbour L, et al. Exposure to anesthetic gases and congenital anomalies in offspring of female registered nurses. *Am J Ind Med*. 2011;54(2):118-27.
 5. Casale T, Caciari T, Rosati MV, et al. Anesthetic gases and occupationally exposed workers. *Environ Toxicol Pharmacol*. 2014;37(1):267-74.
 6. Jafari A, Jafari F, Mohebbi I. Effects of occupational exposure to trace levels of halogenated anesthetics on the liver, kidney, and oxidative stress parameters in operating room personnel. *Toxin Reviews*. 2018:1-10.
 7. Scapellato ML, Mastrangelo G, Fedeli U, et al. A longitudinal study for investigating the exposure level of anesthetics that impairs neurobehavioral performance. *Neurotoxicology*. 2008;29(1):116-23.
 8. Braz MG, Silva MA, Figueiredo DB, et al. Genetic instability assessed by telomere length and micronucleus in physicians exposed to anesthetics. *Environ Mol Mutagen*. 2020;61(8):843-7.
 9. Çakmak G, Eraydın D, Berkkan A, et al. Genetic damage of operating and recovery room personnel occupationally exposed to waste anaesthetic gases. *Hum Exp Toxicol*. 2018:1-8.
 10. Paes ERdC, Braz MG, Lima JTd, et al. DNA damage and antioxidant status in medical residents occupationally exposed to waste anesthetic gases. *Acta Cir Bras*. 2014;29(4):280-6.
 11. Nagella AB, Ravishankar M, Kumar VH. Anaesthesia practice and reproductive outcomes: Facts unveiled. *Indian J Anaesth*. 2015;59(11):706.
 12. Sivaci R, Kahraman A, Serteser M, et al. Cytotoxic effects of volatile anesthetics with free radicals undergoing laparoscopic surgery. *Clin Biochem*. 2006;39(3):293-8.
 13. Lee Y-M, Song BC, Yeum K-J. Impact of volatile anesthetics on oxidative stress and inflammation. *Biomed Res Int*. 2015;1-9.
 14. Malekirad AA, Ranjbar A, Rahzani K, et al. Oxidative stress in operating room personnel: occupational exposure to anesthetic gases. *Hum Exp Toxicol*. 2005;24(11):597-601.
 15. Soleimani E, Moghadam RH, Ranjbar A. Occupational exposure to chemicals and oxidative toxic stress. *Toxicology and Environmental Health Sciences*. 2015;7(1):1-24.
 16. Lucio L, Braz MG, Nascimento Junior Pd, et al. Occupational hazards, DNA damage, and oxidative stress on exposure to waste anesthetic gases. *Rev Bras Anesthesiol*. 2018;68(1):33-41.
 17. Hua H-X, Deng H-B, Huang X-L, et al. Effects of Occupational Exposure to Waste Anesthetic Gas on Oxidative Stress and DNA Damage. *Oxid Med Cell Longev*. 2021:1-8.
 18. Wrońska-Nofer T, Nofer J-R, Jajte J, et al. Oxidative DNA damage and oxidative stress in subjects occupationally exposed to nitrous oxide (N₂O). *Mutat Res*. 2012;731(1):58-63.
 19. Fenech M, Aitken C, Rinaldi J. Folate, vitamin B12, homocysteine status and DNA damage in young Australian adults. *Carcinogenesis*. 1998;19(7):1163-71.
 20. Izdes S, Sardas S, Kadioglu E, et al. DNA damage, glutathione, and total antioxidant capacity in anesthesia nurses. *Arch Environ Occup Health*. 2010;65(4):211-7.
 21. National Institute of Occupational Safety and Health (NIOSH), Criteria for a Recommended Standard: Occupational Exposure to Waste Anesthetic Gases and Vapors. Washington, DC: United States Department of Health, Education, and Welfare (DHEW). 1977; pp. 77–140.
 22. American Conference of Governmental Industrial Hygienists (ACGIH) TLVs and BELs.

- Threshold Limits Values for Chemical Substances and Physical Agents and Biological Exposure Indices. 2021; Cincinnati: ACGIH.
23. Kargar Shouroki F, Neghab M, Mozdarani H, et al. Genotoxicity of inhalational anesthetics and its relationship with the polymorphisms of GSTT1, GSTM1, and GSTP1 genes. *Environ Sci Pollut Res*. 2019;26:3530-41.
 24. National Institute of Occupational Safety and Health (NIOSH) Manual of Analytical Methods, Method 6600. 1994; Atlanta: Centers for Disease Control and Prevention.
 25. Souza KM, Braz LG, Nogueira FR, et al. Occupational exposure to anesthetics leads to genomic instability, cytotoxicity and proliferative changes. *Mutat Res*. 2016;791:42-8.
 26. Braz MG, Souza KM, Lucio LM, et al. Detrimental effects detected in exfoliated buccal cells from anesthesiology medical residents occupationally exposed to inhalation anesthetics: An observational study. *Mutat Res Genet Toxicol Environ Mutagen*. 2018;832-833:61-4.
 27. Wiesner G, Hoerauf K, Schroegendorfer K, et al. High-level, but not low-level, occupational exposure to inhaled anesthetics is associated with genotoxicity in the micronucleus assay. *Anesth Analg*. 2001;92(1):118-22.
 28. Shouroki FK, Neghab M, Mozdarani H, et al. Genotoxicity of inhalational anesthetics and its relationship with the polymorphisms of GSTT1, GSTM1, and GSTP1 genes. *Environ Sci Pollut Res*. 2019;26(4):3530-41.
 29. Meli R, Monnolo A, Annunziata C, et al. Oxidative stress and BPA toxicity: an antioxidant approach for male and female reproductive dysfunction. *Antioxidants*. 2020;9(5):405.
 30. Tenkorang MA, Snyder B, Cunningham RL. Sex-related differences in oxidative stress and neurodegeneration. *Steroids*. 2018;133:21-7.
 31. Viña J, Borrás C, Gambini J, et al. Why females live longer than males? Importance of the upregulation of longevity-associated genes by oestrogenic compounds. *FEBS letters*. 2005;579(12):2541-5.
 32. Türkan H, Aydin A, Sayal A. Effect of volatile anesthetics on oxidative stress due to occupational exposure. *World J Surg*. 2005;29(4):540-2.
 33. Cerit N, Onuk AA, Ellidag HY, et al. Arylesterase and oxidative stress in operating room personnel. *Adv Clin Exp Med*. 2014;23(1):49-55.
 34. Cegin MB, Aslan M, Goktas U, et al. Serum myeloperoxidase (MPO) activity, oxidative and antioxidative parameters in operating room personnel. *J Pak Med Assoc*. 2016;66(666).
 35. Baysal Z, Cengiz M, Ozgonul A, et al. Oxidative status and DNA damage in operating room personnel. *Clin Biochem*. 2009;42(3):189-93.
 36. Bonassi S, Coskun E, Ceppi M, et al. The HUman MicroNucleus project on exfoLiated buccal cells (HUMN XL): the role of life-style, host factors, occupational exposures, health status, and assay protocol. *Mutat Res*. 2011;728(3):88-97.
 37. Sardas S, Izdes S, Ozcagli E, et al. The role of antioxidant supplementation in occupational exposure to waste anaesthetic gases. *Int Arch Occup Environ Health*. 2006;80(2):154-9.
 38. Holt EM, Steffen LM, Moran A, et al. Fruit and vegetable consumption and its relation to markers of inflammation and oxidative stress in adolescents. *American Dietetic Association*. 2009;109(3):414-21.
 39. Ranjbar A, Ghaseminejad S, Takalu H, et al. Anti oxidative stress potential of cinnamon (*cinnamomum zeylanicum*) in operating room personnel; a before/after cross sectional clinical trial. *Int J Pharmacol*. 2007;3(6):482-86.
 40. Sami G, Khoshraftar E, Shayesteh H, et al. The effects of Chamomile tea on antioxidative biomarkers in operating room staff. *Journal of HerbMed Pharmacology*. 2015;4.