

Wastewater-Based Epidemiology for Infectious Disease Surveillance: A Systematic Review and Meta-Analysis Focused on SARS-Cov-2, with Narrative Synthesis for Other Pathogens

Soleiman Forouzandeh^{1,2}, Amin Salehi-Abargouei³, Ali Asghar Ebrahimi^{1*}

¹ Environmental Sciences and Technology Research Center, School of Public Health, Shahid Sadoughi University of Medical Sciences, Yazd, Iran.

² Department of Environmental Health Engineering, School of Health, Shahrekord University of Medical Sciences, Shahrekord, Iran.

³ Department of Nutrition, School of Public Health, Faculty of Health, Shahid Sadoughi University of Medical Sciences, P O Box 8915173160, Yazd, Iran.

ARTICLE INFO

REVIEW ARTICLE

Article History:

Received: 17 May 2025

Accepted: 20 July 2025

*Corresponding Author:

Ali Asghar Ebrahimi

Email:

ebrahimi20007@gmail.com

Tel:

+98 913 2679641

Keywords:

Sewage; COVID-19;

SARS-Cov-2;

Environmental Monitoring;

Systematic Reviews As Topic;

Meta-Analysis As Topic;

Public Health Surveillance.

ABSTRACT

Introduction: Wastewater-based epidemiology (WBE) has emerged as a valuable approach for environmental management and public health surveillance. By detecting viral RNA and biomarkers in wastewater, WBE provides community-level early warning signals that can support outbreak preparedness and guide policy-making.

Methods: We conducted a systematic review and meta-analysis according to the PRISMA 2020 guidelines. Eligible studies published until March 2025 were screened from PubMed, Web of Science, and Scopus. Data on study design, sample type, detection methods, positivity rates, and temporal associations with clinical indicators were extracted. Pooled positivity rates were calculated using random-effects models, heterogeneity was assessed, and subgroup analyses were conducted.

Results: Twenty-nine studies on SARS-CoV-2 wastewater surveillance were included in this review. The pooled positivity rate was 59.5% (95% CI: 49.6–68.7), with signals detected earliest in the sludge samples (98.8%). Wastewater indicators preceded reported clinical cases by a median of six days. Evidence has also demonstrated WBE's applicability of WBE for influenza, RSV, norovirus, polio, and antimicrobial resistance, reinforcing its value beyond COVID-19.

Citation: Forouzandeh S, Salehi-Abargouei A, Ebrahimi AA. *Wastewater-Based Epidemiology for Infectious Disease Surveillance: A Systematic Review and Meta-Analysis Focused on SARS-Cov-2, with Narrative Synthesis for Other Pathogens*. J Environ Health Sustain Dev. 2025; 10(3): 2712-24.

Introduction

Wastewater-based epidemiology (WBE) involves the detection of pathogens and biomarkers in sewage to monitor community health. This approach gained global attention during the COVID-19 pandemic as a cost-effective,

non-invasive surveillance tool that can detect outbreaks earlier than clinical testing¹⁻³. WBE has since expanded to track influenza, respiratory syncytial virus (RSV), monkeypox, polio, and antimicrobial resistance genes⁴⁻¹⁵.

The primary rationale for WBE lies in its ability

to capture signals from symptomatic, pre-symptomatic, and asymptomatic individuals, thus providing an integrated population-level signal¹⁶⁻¹⁹. Wastewater surveillance has been implemented at the national level in several countries, including the U.S. National Wastewater Surveillance System²⁰ and WHO Environmental Surveillance Initiatives²¹.

Despite the rapid growth in WBE research, uncertainties remain regarding pooled positivity rates across study settings, methodological heterogeneity, and the consistency of lead-time advantages relative to clinical surveillance. Furthermore, the role of WBE in pathogens beyond SARS-CoV-2 is less systematically documented.

This systematic review and meta-analysis aimed to:

- Quantification of pooled positivity rates of SARS-CoV-2 in wastewater samples.
- Assess the lead time between wastewater signals and clinical case reports.
- Evaluation of the sources of heterogeneity (sample matrices and study quality).
- Synthesize emerging evidence for the WBE of influenza, RSV, polio, and other pathogens.

By addressing these objectives, this study provides evidence to support the integration of WBE into global infectious disease surveillance systems for pandemic preparedness and beyond.

The innovation of this study lies in providing the most comprehensive synthesis to date, covering evidence up to March 2025, while simultaneously performing a quantitative meta-analysis of SARS-CoV-2 and a narrative synthesis of other pathogens. By combining both statistical and qualitative approaches, this study not only clarifies the pooled detection rates and lead-time advantages, but also highlights the broader applicability of WBE as a multi-pathogen

surveillance tool. This dual focus distinguishes the present study from previous reviews and underscores its importance for sustainable public health preparedness.

Methods

Search strategy and selection criteria

We conducted a systematic review following the PRISMA 2020 guidelines²². Searches were performed in PubMed, Web of Science, and Scopus through March 2025 using combinations of keywords: wastewater, sewage, wastewater-based epidemiology, surveillance, SARS-CoV-2, COVID-19, influenza, RSV, norovirus, adenovirus, polio, monkeypox, and antimicrobial resistance. No language restrictions were imposed. The reference lists of relevant reviews were hand-searched to identify additional studies. The detailed search strategy for each database is provided in Table 1. The review protocol was not prospectively registered in PROSPERO or any other international registries.

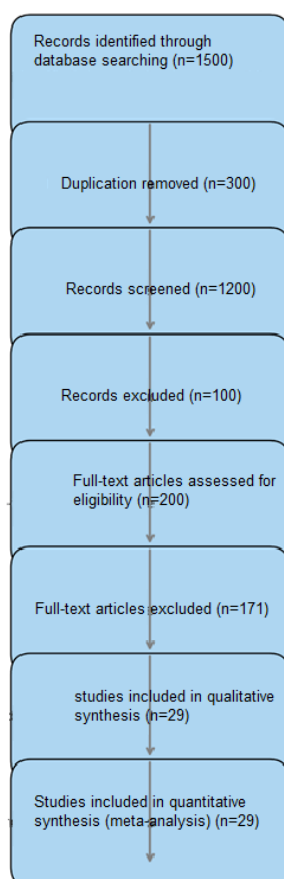
Studies were eligible if they met the following criteria:

- Reported wastewater detection of SARS-CoV-2 and other pathogens
- Included at least 10 samples or >1 week of monitoring
- Reported positivity rates and/or quantitative viral load data
- Linked wastewater data to epidemiological indicators, where available.

The exclusion criteria were modeling-only studies without empirical wastewater data, case reports, and conference abstracts. The study selection process is summarized in a PRISMA 2020 flow diagram (Figure 1), which shows the number of records identified, screened, excluded (with reasons), and included in the final review.

Table 1: Search strategies applied in electronic databases

Database	Search strategy	Coverage / Notes
PubMed	("wastewater" OR "sewage" OR "wastewater-based epidemiology" OR "wastewater surveillance") AND ("SARS-CoV-2" OR "COVID-19" OR "influenza" OR "RSV" OR "norovirus" OR "adenovirus" OR "poliovirus" OR "monkeypox" OR "antimicrobial resistance")	Up to March 2025; no language restriction
Web of Science	TS=("wastewater" OR "sewage" OR "wastewater-based epidemiology" OR "wastewater surveillance") AND TS=("SARS-CoV-2" OR "COVID-19" OR "influenza" OR "RSV" OR "norovirus" OR "adenovirus" OR "poliovirus" OR "monkeypox" OR "antimicrobial resistance")	Up to March 2025; no language restriction
Scopus	TITLE-ABS-KEY("wastewater" OR "sewage" OR "wastewater-based epidemiology" OR "wastewater surveillance") AND TITLE-ABS-KEY("SARS-CoV-2" OR "COVID-19" OR "influenza" OR "RSV" OR "norovirus" OR "adenovirus" OR "poliovirus" OR "monkeypox" OR "antimicrobial resistance")	Up to March 2025; no language restriction

**Figure 1:** PRISMA 2020 flow diagram for study selection.**Data extraction**

Two reviewers independently extracted the data into standardized forms (Supplementary Tables S3 and S4, Supplementary Material). The extracted variables included study setting, sample matrix (raw wastewater, treated effluent, sludge, hospital wastewater), concentration and extraction methods, target gene(s), positivity rate, lead time relative to

clinical cases, and quality assessment score. Discrepancies were resolved by consensus agreement.

Risk of bias and quality assessment

Study quality was assessed using a modified checklist adapted from Corchis-Scott et al.^{23, 24}. The criteria included clarity of objectives,

sampling strategy, analytical methods, and reporting of clinical data linkage. The quality scores are summarized in Table S1.

Statistical analysis

The primary outcome was the pooled proportion of positive wastewater samples. A random-effects meta-analysis was conducted using the DerSimonian and Laird method²⁵. Proportions were logit-transformed and back-transformed for reporting purposes. Heterogeneity was assessed using Cochran's Q and I² statistics²⁶. Publication bias was evaluated using funnel plots and Egger's test²⁷. Subgroup analyses were performed based on the sample matrix (Figure 4) and study quality (Figure 5). Sensitivity analyses restricted the dataset to studies with quality scores >50. The temporal lead-time was synthesized as the median and interquartile range across the eligible studies

(Figure 6).

All statistical analyses were performed using R (version 4.2.0) with the meta and metafor packages.

Results

A total of 29 eligible studies on SARS-CoV-2 wastewater surveillance were included in the meta-analysis (Figure 1); the complete study list is provided in Supplementary Tables S2 and S3. The key characteristics of the included studies, including the country, study period, sample type, and detection methods, are summarized in Table 2. The pooled positivity rate across all matrices was 59.5% (95% CI: 49.6–68.7; Table S1; Figure 2), with substantial heterogeneity (I² = 93.5%). Funnel plot inspection showed approximate symmetry (Figure 3), and Egger's regression test did not indicate a significant publication bias²⁷.

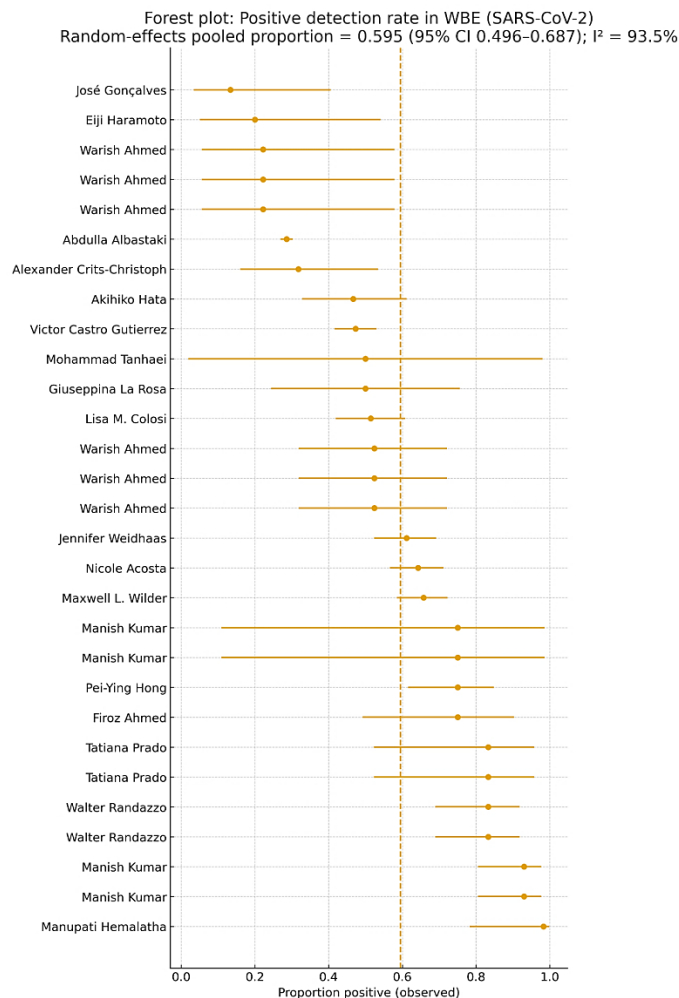


Figure 2: Forest plot: Proportion of SARS-CoV-2–positive wastewater samples.

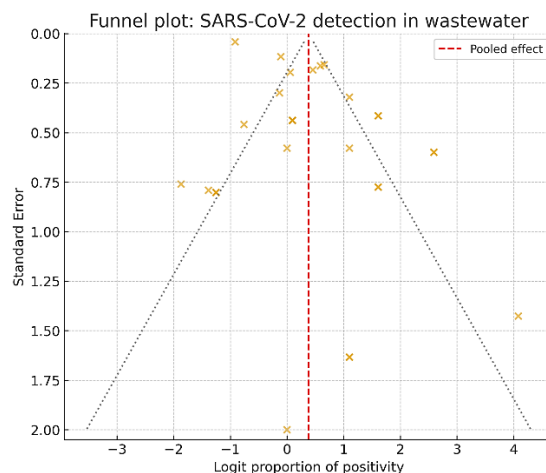


Figure 3: Funnel plot—Publication bias assessment of the primary outcome.

Table 2: Characteristics of the studies included in the systematic review and meta-analysis.

Author & ref (Year)	Country	Sample type	Detection method	Positive / Total (%)
Albastaki ²⁸ (2020)	UAE – Dubai	Wastewater (grab)	RT-qPCR	829 / 2900 (28.6)
Hata (2020) ²⁹	Japan – Ishikawa/Toyama	Influent wastewater (grab)	RT-PCR	21 / 45 (46.7)
Crits-Christoph (2020) ³⁰	USA – California	Wastewater (24h composite)	RT-qPCR	7 / 22 (31.8)
Haramoto (2020) ³¹	Japan – Yamanashi	Wastewater & river water (grab)	Nested PCR/qPCR	2 / 10 (20.0)
Ahmed (2020) ^{32, 33}	Bangladesh – Noakhali	Untreated wastewater	RT-PCR	12 / 16 (75.0)
La Rosa (2020) ³⁴	Italy – Milan/Rome	Raw wastewater (24h composite)	Nested RT- PCR/qPCR	6 / 12 (50.0)
Weidhaas (2020) ³⁵	USA – Utah	Wastewater (composite & grab)	RT-qPCR	77 / 126 (61.1)
Gonçalves (2020) ³⁶	Slovenia – Ljubljana	Hospital wastewater (untreated)	RT-qPCR	2 / 15 (13.3)
Colosi (2020) ³⁷	USA – Virginia	Hospital & dorm wastewater	PCR	54 / 105 (51.4)
Kumar (2020) ³⁸	India – Gandhinagar	Influent wastewater (grab)	RT-PCR	40 / 43 (93.0)
Kumar (2020) ³⁹	India – Ahmedabad	Influent/effluent (composite)	RT-PCR	1.5 / 2 (75.0)
Hemalatha (2020) ⁴⁰	India – Hyderabad	Raw & treated wastewater	RT-PCR	29.5 / 30 (98.3)
Wilder (2020) ⁴¹	USA – New York (Upstate)	Wastewater (24h composite)	RT-qPCR	119 / 181 (65.7)
Tanhaei (2020) ⁴²	Iran – Tehran	Untreated & treated wastewater	RT-qPCR	0.5 / 1 (50.0)
Acosta (2020) ⁴³	Canada – Calgary	Untreated wastewater (composite)	RT-qPCR + sequencing	106 / 165 (64.2)
Hong (2020) ⁴⁴	Saudi Arabia – Jeddah	Hospital wastewater (grab)	RT-qPCR	39 / 52 (75.0)
Prado (2020) ⁴⁵	Brazil – Niterói	Wastewater (10h composite)	RT-qPCR	10 / 12 (83.3)
Castro-Gutierrez (2020) ⁴⁶	England	Wastewater (time- proportional)	RT-qPCR	140 / 296 (47.3)
Randazzo (2020) ⁴⁷	Spain – Murcia	Influent/effluent (grab)	RT-qPCR	35 / 42 (83.3)
Ahmed (2020) ³²	Australia – Queensland	Untreated wastewater (grab)	RT-qPCR	2 / 9 (22.2)
Ahmed (2020) ⁴⁸	Australia	Wastewater	RT-qPCR	11 / 21 (52.4)

Subgroup analysis

The detection rates varied significantly according to the matrix (Figure 4). Viral RNA was most frequently detected in sludge samples (98.8%), followed by raw wastewater (53.6%), and

untreated wastewater influent (51.2%). Hospital wastewater samples had a lower positivity rate (33.1%). These findings are consistent with earlier single-site studies that reported higher recovery in solids than in liquid fractions^{1, 3, 49, 50}.

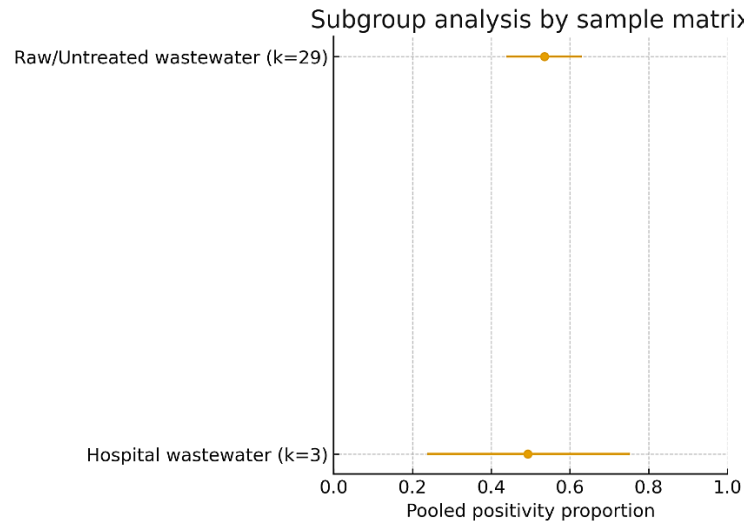


Figure 4: Forest plot—Subgroup analyses by sample matrix.

Sensitivity analysis

Limiting the analysis to higher-quality studies (quality score >50; **Table S2**) yielded a pooled positivity rate of 59.7% (95% CI: 48.4–70.2; **Figure 5**). Heterogeneity decreased modestly ($I^2 = 78.2\%$), confirming the robustness of the main findings.

Temporal lead-time

In 15 studies that compared wastewater signals with clinical case data, wastewater RNA preceded reported cases by a median of six days (IQR: 2–19.5; Table 4 and Figure 6). Several studies have reported lead times exceeding two weeks⁵⁰⁻⁵².

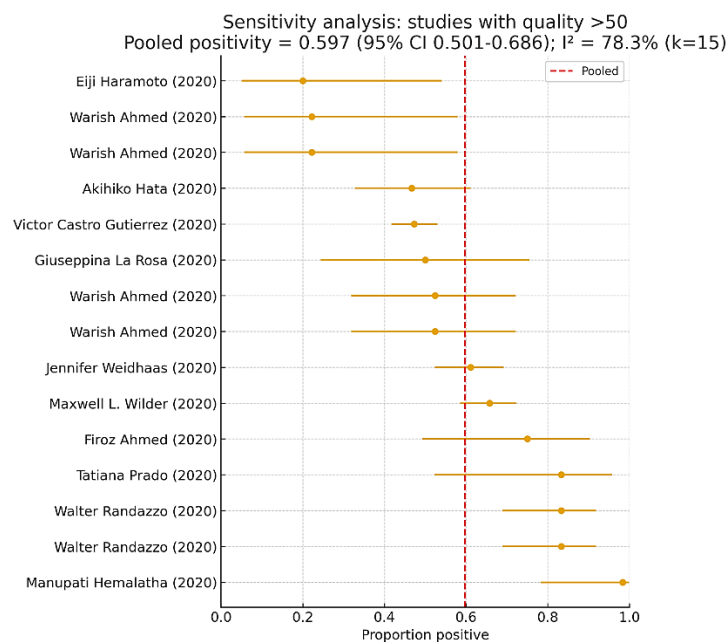


Figure 5: Sensitivity analysis—Studies with quality score > 50.

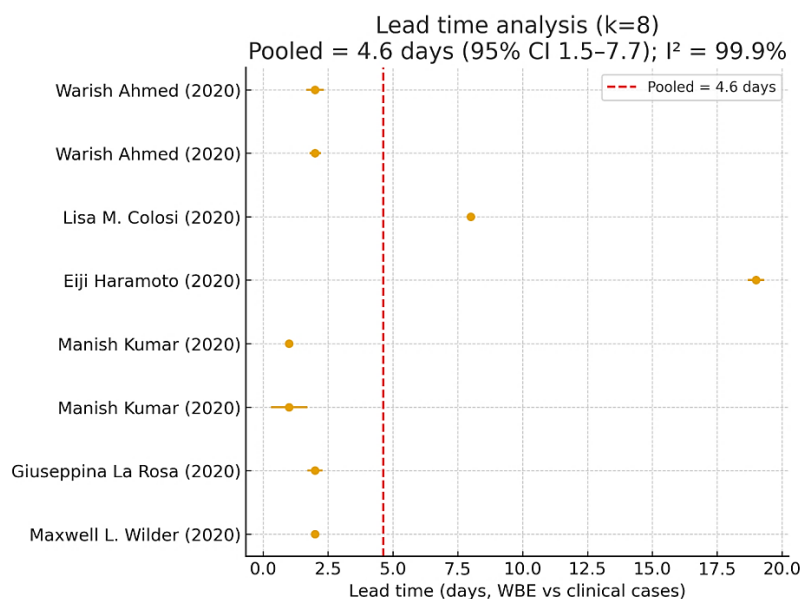


Figure 6: Forest plot—Lead time (days) of wastewater signal vs. clinical cases.

Table 3: Summary of lead-time estimates (days) between wastewater signals and clinical cases across the included studies.

Author& ref	Year	Country	Lead time (days)	Sample size (if reported)
Warish Ahmed ³²	2020	Australia- Brisbane	2.0	9.0
Warish Ahmed ⁵³	2020	Australia- Brisbane	2.0	21.0
Gemma Chavarria-Miró ⁵⁴	2020	Spain-Barcelona	41.0	
Lisa M. Colosi ³⁷	2020	USA-Virginia	8.0	105.0
PatrickM. D'Aoust ⁵⁵	2020	Canada-Ottawa	2.0	
Gislaine Fongaro ⁵⁶	2019-2020	Brazil-Florianopolis	60.0	
Eiji Haramoto ⁵⁷	2020	Japan-Yamanashi	19.0	10.0
Kouichi Kitamura ⁵⁸	2020	Japan	19.0	
Manish Kumar ⁵⁹	2020	India-Gandhinagar	1.0	43.0
Manish Kumar ⁵⁹	2020	India-Gandhinagar	1.0	2.0
Giuseppina La Rosa ⁶⁰	2020	Italy-Milan and Rome	2.0	12.0
Javier Martin ⁶¹	2020	South-East England	3.0	
Gertjan Medema ⁶²	2020	Netherlands-Amsterdam Den Haag Utrecht Apeldoorn Amersfoort Schiphol	6.0	
Richard G. Melvin ⁶³	2020	USA-Minnesota	20.0	
Jordan Peccia ⁶⁴	2020	USA-New Haven	7.0	
Maxwell L. Wilder ⁴¹	2020	USA-Upstate New York	2.0	181.0
Gislaine Fongaro ⁶⁵	2020	Brazil-Florianopolis, Santa Catarina	97.0	
Jordan Peccia ⁶⁴	2020	USA-New Haven, Connecticut	2.0	

Other pathogens

Emerging evidence has shown the applicability of WBE to pathogens beyond SARS-CoV-2. Influenza A and B were consistently detected in wastewater during 2022–2023⁴, with pooled positivity of 36% for influenza B and 62% for influenza A¹⁰. RSV RNA has been detected in multiple community-level studies⁴, and

monkeypox DNA was reported in wastewater during the 2022 European outbreaks¹¹. Surveillance for norovirus and adenovirus has long been established^{66, 67}, and poliovirus monitoring remains integral to global eradication efforts¹⁴. Finally, antimicrobial resistance genes are increasingly tracked in wastewater, highlighting WBE's role of WBE in One Health

surveillance^{17, 18, 68}.

Discussion

This meta-analysis confirms that WBE is a robust tool for the monitoring of infectious diseases. The pooled positivity for SARS-CoV-2 (~60%) was consistent with previous reviews²³ and observational studies in Europe, Asia, and North America¹⁻³. Temporal analysis showed that wastewater signals preceded clinical data by nearly one week on average, echoing prior modeling studies estimating lead times of 5–11 days^{50, 69}.

The subgroup findings highlighted the methodological determinants of positivity. Higher detection rates in sludge than in influent wastewater are consistent with viral partitioning to solids^{3, 49}. Hospital effluent showed lower positivity, possibly due to smaller, less representative populations. Methodological heterogeneity—sample type, concentration method, and RT-qPCR assays—explains part of the variation, as reflected in the high I^2 values. The need for methodological harmonization is widely recognized^{19, 70}.

Beyond SARS-CoV-2, wastewater surveillance for influenza, RSV, and other pathogens demonstrates WBE's versatility. Recent reviews^{10, 66} have emphasized the promise of multiplex surveillance platforms. The detection of monkeypox¹¹ and ongoing poliovirus monitoring¹⁴ illustrate WBE's utility for emerging and re-emerging infections. Moreover, the monitoring of antimicrobial resistance genes positions WBE as an essential One Health tool¹⁷.

The strengths of this study include its comprehensive coverage through March 2025, the use of a quantitative meta-analysis combined with a narrative synthesis, and the application of a structured quality assessment tool. This dual approach provides both statistical rigor and a broader contextual understanding, offering policymakers practical insights into the implementation of WBE.

The limitations of this study must also be acknowledged. High heterogeneity persisted despite subgroup analyses, reflecting differences in the study design, population coverage, and laboratory

methods. In addition, studies from low- and middle-income countries remain underrepresented, limiting the generalizability of the findings to global contexts. Reliance on observational designs with variable clinical data linkage also restricts causal interpretation.

In comparison with earlier systematic reviews^{71, 72}, this study extends the evidence base by incorporating data through 2025, providing updated pooled positivity estimates, and uniquely synthesizing the findings for non-SARS-CoV-2 pathogens. This broader perspective highlights the evolving role of WBE, from a COVID-19-focused tool to a cornerstone of multi-pathogen surveillance systems.

The public health implications are clear: WBE provides a cost-effective, scalable, and non-invasive means of surveillance. The integration of WBE into national programs, such as the CDC National Wastewater Surveillance System²⁰ and WHO guidance²¹, represents a pathway toward global pandemic preparedness.

Conclusion

This systematic review and meta-analysis addressed four primary objectives: quantifying the pooled positivity rates of SARS-CoV-2 in wastewater, assessing the lead time between wastewater signals and reported clinical cases, evaluating the sources of methodological heterogeneity, and synthesizing evidence for pathogens beyond SARS-CoV-2. Our findings confirmed a pooled positivity rate of approximately 60%, with the highest detection in sludge samples, and demonstrated that wastewater signals preceded clinical cases by nearly one week. Methodological variability, including differences in sample matrices and RT-qPCR protocols, explains much of the heterogeneity.

Beyond COVID-19, this review synthesizes emerging data on influenza, RSV, norovirus, poliovirus, and antimicrobial resistance, highlighting WBE's versatility as a multi-pathogen surveillance tool. These results demonstrate that WBE provides not only biomedical signals but also an integrated environmental

management framework for public health surveillance.

Strengthening methodological standardization, including sampling strategies, concentration methods, and normalization approaches, remains essential for comparability and policy integration. By linking environmental monitoring to health system preparedness, this study contributes directly to sustainable development and to pandemic resilience.

Acknowledgments

The authors would like to thank the Department of Environmental Health Engineering at Shahid Sadoughi University of Medical Sciences in Yazd (Iran) for obtaining the latest literature and information on Covid 19.

Conflict of Interest

There are no conflicts to declare

Funding

This project is Funded by Shahid Sadoughi University of Medical Sciences, Yazd, Iran

Ethical Considerations

This study is a systematic review of previously published literature and does not involve human participants or animals. Therefore, ethical approval and informed consent were not required.

Code of Ethics

The authors affirm that the study was conducted in accordance with ethical principles of research integrity and reporting. No ethical approval code was necessary due to the nature of the study.

Authors' Contributions

Soleiman Forouzandeh contributed to the conception and design of the study, data acquisition, and initial drafting of the manuscript.

Amin Salehi Abargouei contributed to data analysis, interpretation of the findings, and manuscript revisions.

Ali Asghar Ebrahimi supervised the project, provided critical revisions, and is the corresponding author.

All authors read and approved the final manuscript

Appendix A. Supplementary data

Supplementary data to this article can be found online at:
https://jehsd.ssu.ac.ir/files/site1/files/supplements_soleiman_forouzandeh.pdf?grand=2824

This is an Open-Access article distributed in accordance with the terms of the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt, and build upon this work for commercial use.

References

1. Peccia J, Zulli A, Brackney DE, et al. Measurement of SARS-CoV-2 RNA in wastewater tracks community infection dynamics. *Nat Biotechnol.* 2020;38(10):1164-7.
2. Ahmed W, Angel N, Edson J, et al. First confirmed detection of SARS-CoV-2 in untreated wastewater in Australia: a proof of concept for the wastewater surveillance of COVID-19 in the community. *Sci Total Environ.* 2020;728:138764.
3. Wu F, Zhang J, Xiao A, et al. SARS-CoV-2 titers in wastewater are higher than expected from clinically confirmed cases. *mSystems.* 2021;6(1):10-128.
4. Maida CM, Mazzucco W, Priano W, et al. Detection of influenza virus in urban wastewater during the season 2022/2023 in Sicily, Italy. *Front Public Health.* 2024;12:1383536.
5. Carducci A, Federigi I, Pagani A, et al. Wastewater-based surveillance of respiratory viruses in Northern Tuscany (Italy): challenges and added value for public health purposes. *Sci Total Environ.* 2024;957:177752.
6. Allen DM, Reyne MI, Allingham P, et al. Genomic analysis and surveillance of respiratory syncytial virus using wastewater-based epidemiology. *J Infect Dis.* 2024;230(4):e895-e904.
7. Lämsivaara A, Lehto K-M, Hyder R, et al. Wastewater-based surveillance of respiratory syncytial virus epidemic at the national level in Finland. *ACS ES T Water.* 2024;4(6):2403-11.
8. Beach M, Corchis-Scott R, Geng Q, et al.

- Wastewater-based surveillance of respiratory syncytial virus reveals a temporal disconnect in disease trajectory across an active international land border. *Environment & Health*. 2025;3(4):425-35.
9. Zhang Z, Li Q, He F, et al. Sewage surveillance revealed the seasonality and prevalence of respiratory syncytial virus and its implications for seasonal immunization strategy in low and middle-income regions of China. *Water Res*. 2025;270:122828.
 10. Viviani L, Vecchio R, Pariani E, et al. Wastewater-based epidemiology of influenza viruses: a systematic review. *Sci Total Environ*. 2025;986:179706.
 11. Islam MA, Kumar R, Sharma P, et al. Wastewater-based surveillance of mpox (Monkeypox): an early surveillance tool for detecting hotspots. *Curr Pollut Rep*. 2024;10(2):312-25.
 12. Calabria de Araujo J, Carvalho A, Leal C, et al. Detection of Mpox Virus Using a Wastewater Surveillance Approach in Brazil. 2024.
 13. Lee JH, Sim G, Park HJ, et al. Monkeypox virus detection using wastewater surveillance during the Mpox outbreak in the Republic of Korea. *J Environ Chem Eng*. 2024;12(6):114775.
 14. Böttcher S, Kreibich J, Wilton T, et al. Detection of circulating vaccine-derived poliovirus type 2 (cVDPV2) in wastewater samples: a wake-up call, Finland, Germany, Poland, Spain, the United Kingdom, 2024. *Eurosurveillance*. 2025;30(3):2500037.
 15. Wiecek M, Gad B, Krzysztozek A, et al. Detection of vaccine-derived poliovirus type 2 from sewage samples and public health response, Poland, November to December 2024. *Eurosurveillance*. 2025;30(1):2400805.
 16. Silvester R, Perry WB, Webster G, et al. Metagenomic profiling of hospital wastewater: a comprehensive national scale analysis of antimicrobial resistance genes and opportunistic pathogens. *J Infect*. 2025:106503.
 17. Lan L, Wang Y, Chen Y, et al. A review on the prevalence and treatment of antibiotic resistance genes in hospital wastewater. *Toxics*. 2025;13(4):263.
 18. Hotor P, Kotey FC, Donkor ES. Antibiotic resistance in hospital wastewater in West Africa: a systematic review and meta-analysis. *BMC Public Health*. 2025;25(1):1364.
 19. Kitajima M, Sassi HP, Torrey JR. Pepper mild mottle virus as a water quality indicator. *NPJ Clean Water*. 2018;1(1):19.
 20. Adams C, Bias M, Welsh RM, et al. The national wastewater surveillance system (NWSS): from inception to widespread coverage, 2020–2022, United States. *Sci Total Environ*. 2024;924:171566.
 21. Organization WH. Environmental surveillance for SARS-CoV-2 to complement other public health surveillance: World Health Organization; 2023.
 22. Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. 2021;372.
 23. Rogers E. Variability and uncertainty in SARS-CoV-2 wastewater-based surveillance normalization: a systematic review: University of Washington; 2025.
 24. Corchis-Scott R, Geng Q, Seth R, et al. Averting an outbreak of SARS-CoV-2 in a university residence hall through wastewater surveillance. *Microbiol Spectr*. 2021;9(2):e00792-21.
 25. DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials*. 1986;7(3):177-88.
 26. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Statistics in Medicine*. 2002;21(11):1539-58.
 27. Egger M, Smith GD, Schneider M, et al. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315(7109):629-34.
 28. Albastaki A, Naji M, Lootah R, et al. First confirmed detection of SARS-COV-2 in untreated municipal and aircraft wastewater in Dubai, UAE: the use of wastewater based epidemiology as an early warning tool to

- monitor the prevalence of COVID-19. *Sci Total Environ.* 2021;760:143350.
29. Hata A, Honda R, Honda R. Potential sensitivity of wastewater monitoring for SARS-CoV-2: comparison with Norovirus Cases. *Environmental Science and Technology.* 2020;54(11):6451-2.
 30. Crits-Christoph A, Kantor RS, Olm MR, et al. Genome sequencing of sewage detects regionally prevalent SARS-CoV-2 variants. *mBio.* 2021;12(1):1-9.
 31. Haramoto E, Malla B, Thakali O, et al. First environmental surveillance for the presence of SARS-CoV-2 RNA in wastewater and river water in Japan. *Sci Total Environ.* 2020;737:140405.
 32. Ahmed W, Angel N, Edson J, et al. First confirmed detection of SARS-CoV-2 in untreated wastewater in Australia: a proof of concept for the wastewater surveillance of COVID-19 in the community. *Sci Total Environ.* 2020;728:138764.
 33. Ahmed F, Islam MA, Kumar M, et al. First detection of SARS-CoV-2 genetic material in the vicinity of COVID-19 isolation Centre in Bangladesh: variation along the sewer network. *Sci Total Environ.* 2021;776:145724.
 34. La Rosa G, Iaconelli M, Mancini P, et al. First detection of SARS-CoV-2 in untreated wastewaters in Italy. *Sci Total Environ.* 2020;736:139652.
 35. Weidhaas J, Aanderud ZT, Roper DK, et al. Correlation of SARS-CoV-2 RNA in wastewater with COVID-19 disease burden in sewersheds. *Sci Total Environ.* 2021;775:145790.
 36. Goncalves J, Koritnik T, Mioc V, et al. Detection of SARS-CoV-2 RNA in hospital wastewater from a low COVID-19 disease prevalence area. *Sci Total Environ.* 2021;755.
 37. Colosi LM, Barry KE, Kotay SM, et al. Development of wastewater pooled surveillance of SARS-CoV-2 from congregate living settings. *Appl Environ Microbiol.* 2020:2020-10.
 38. Kumar M, Mohapatra S, Mazumder P, et al. Making waves perspectives of modelling and monitoring of SARS-CoV-2 in aquatic environment for COVID-19 pandemic. *Curr Pollut Rep.* 2020:1-12.
 39. Kumar M, Patel AK, Shah AV, et al. First proof of the capability of wastewater surveillance for COVID-19 in India through detection of genetic material of SARS-CoV-2. *Sci Total Environ.* 2020;746:141326.
 40. Hemalatha M, Kiran U, Kuncha SK, et al. Surveillance of SARS-CoV-2 spread using wastewater-based epidemiology: comprehensive study. *Sci Total Environ.* 2021;768:144704.
 41. Wilder ML, Middleton F, Larsen DA, et al. Co-quantification of crAssphage increases confidence in wastewater-based epidemiology for SARS-CoV-2 in low prevalence areas. *Water Res X.* 2021;11:100100.
 42. Tanhaei M, Mohebbi SR, Hosseini SM, et al. The first detection of SARS-CoV-2 RNA in the wastewater of Tehran, Iran. *Environmental Science and Pollution Research.* 2021:1-8.
 43. Acosta N, Bautista M, Hollman J, et al. Wastewater monitoring of SARS-CoV-2 from acute care hospitals identifies nosocomial transmission and outbreaks. *medRxiv.* 2021:2021-02.
 44. Hong PY, Rachmadi AT, Mantilla-Calderon D, et al. Estimating the minimum number of SARS-CoV-2 infected cases needed to detect viral RNA in wastewater: to what extent of the outbreak can surveillance of wastewater tell us?. *Environ Res.* 2021;195:110748.
 45. Prado T, Fumian TM, Mannarino CF, et al. Wastewater-based epidemiology as a useful tool to track SARS-CoV-2 and support public health policies at municipal level in Brazil. *Water Res.* 2021;191.
 46. Gutierrez VC, Hassard F, Vu M, et al. Monitoring occurrence of SARS-CoV-2 in school populations: a wastewater-based approach. *medRxiv.* 2021:2021-03.
 47. Randazzo W, Truchado P, Cuevas-Ferrando E, et al. SARS-CoV-2 RNA in wastewater anticipated COVID-19 occurrence in a low prevalence area. *Water Res.* 2020;181: 115942.

48. Ahmed W, Bertsch PM, Bibby K, et al. Decay of SARS-CoV-2 and surrogate murine hepatitis virus RNA in untreated wastewater to inform application in wastewater-based epidemiology. *Environ Res.* 2020;191:110092.
49. Rusiñol M, Martínez-Puchol S, Forés E, et al. Concentration methods for the quantification of coronavirus and other potentially pandemic enveloped virus from wastewater. *Curr Opin Environ Sci Health.* 2020;17:21-8.
50. Olesen SW, Imakaev M, Duvallet C. Making waves: defining the lead time of wastewater-based epidemiology for COVID-19. *Water Res.* 2021;202:117433.
51. Kumar M, Jiang G, Thakur AK, et al. Lead time of early warning by wastewater surveillance for COVID-19: geographical variations and impacting factors. *Chemical Engineering Journal.* 2022;441:135936.
52. Bibby K, Bivins A, Wu Z, et al. Making waves: plausible lead time for wastewater based epidemiology as an early warning system for COVID-19. *Water Res.* 2021;202:117438.
53. Ahmed W, Bertsch PM, Angel N, et al. Detection of SARS-CoV-2 RNA in commercial passenger aircraft and cruise ship wastewater: a surveillance tool for assessing the presence of COVID-19 infected travellers. *J Travel Med.* 2021;27(5):1-11.
54. Chavarria-Miró G, Anfruns-Estrada E, Martínez-Velázquez A, et al. Time evolution of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in wastewater during the first pandemic wave of COVID-19 in the Metropolitan Area of Barcelona, Spain. *Appl Environ Microbiol.* 2021;87(7).
55. D'Aoust PM, Mercier E, Montpetit D, et al. Quantitative analysis of SARS-CoV-2 RNA from wastewater solids in communities with low COVID-19 incidence and prevalence. *Water Res.* 2021;188:116560.
56. Fongaro G, Stoco PH, Souza DSM, et al. SARS-CoV-2 in human sewage in Santa Catalina, Brazil, November 2019. *medRxiv.* 2020.
57. Haramoto E, Malla B, Thakali O, et al. First environmental surveillance for the presence of SARS-CoV-2 RNA in wastewater and river water in Japan. *Sci Total Environ.* 2020;737:140405.
58. Kitamura K, Sadamasu K, Muramatsu M, et al. Efficient detection of SARS-CoV-2 RNA in the solid fraction of wastewater. *Sci Total Environ.* 2021;763.
59. Kumar M, Joshi M, Patel AK, et al. Unravelling the early warning capability of wastewater surveillance for COVID-19: a temporal study on SARS-CoV-2 RNA detection and need for the escalation. *Environ Res.* 2021;196.
60. La Rosa G, Iaconelli M, Mancini P, et al. First detection of SARS-CoV-2 in untreated wastewaters in Italy. *Sci Total Environ.* 2020;736.
61. Martin J, Klapsa D, Wilton T, et al. Tracking SARS-CoV-2 in sewage: evidence of changes in virus variant predominance during COVID-19 pandemic. *Viruses.* 2020;12(10).
62. Medema G, Heijnen L, Elsinga G, et al. Presence of SARS-Coronavirus-2 RNA in sewage and correlation with reported COVID-19 prevalence in the early stage of the epidemic in the Netherlands. *Environ Sci Technol Lett.* 2020;7(7):511-6.
63. Melvin RG, Chaudhry N, Georgewill O, et al. Predictive power of SARS-CoV-2 wastewater surveillance for diverse populations across a large geographical range. *medRxiv.* 2021.
64. Peccia J, Zulli A, Brackney DE, et al. Measurement of SARS-CoV-2 RNA in wastewater tracks community infection dynamics. *Nat Biotechnol.* 2020;38(10):1164-7.
65. Fongaro G, Stoco PH, Souza DSM, et al. The presence of SARS-CoV-2 RNA in human sewage in Santa Catarina, Brazil, November 2019. *Sci Total Environ.* 2021;778:146198.
66. Bonanno Ferraro G, Brandtner D, Mancini P, et al. Eight Years of norovirus surveillance in urban wastewater: insights from next-generation. *Viruses.* 2025;17(1):130.
67. Wang Y, Amarasiri M, Oishi W, et al. Aptamer-based biosensors for wastewater

- surveillance of influenza virus, SARS-CoV-2, and norovirus: a comprehensive review. *Water Res.* 2025;123484.
68. Nafai M, Bouhoudan A, Mourabit F, et al. Antibiotic resistance from untreated hospital wastewater discharges: a literature review. *Int J Environ Stud.* 2025;1-24.
69. Dai X, Acosta N, Lu X, et al. A Bayesian framework for modeling COVID-19 case numbers through longitudinal monitoring of SARS- CoV- 2 RNA in wastewater. *Statistics in Medicine.* 2024;43(6):1153-69.
70. Kantor RS, Nelson KL, Greenwald HD, et al. Challenges in measuring the recovery of SARS-CoV-2 from wastewater. *Environ Sci Technol.* 2021;55(6):3514-9.
71. Kilaru P, Hill D, Anderson K, et al. Wastewater surveillance for infectious disease: a systematic review. *Am J Epidemiol.* 2023;192(2):305-22.
72. Carducci A, Gerba CP, Haramoto E, et al. SARS-CoV-2 in wastewater: State of the Knowledge and Research Needs. 2020.